IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:

HEATH, et al.

Serial No.: 08/699,716

Filed: 27 August 1996

For: RECOMBINANT F1-V PLAGUE VACCINE



Art Unit: 1645

Examiner: Duffy, Patricia Ann

Atty. Dckt: 003/029/SAP

AFFIDAVIT OF GEORGE W. ANDERSON, JR.

- 1. I, George W. Anderson, Jr., an inventor of the above-referenced application and resident of Smithsburg, MD, declare the following:
- 2. My curriculum vitae is attached.
- 3. Arthur M. Friedlander, David G. Heath, Susan L. Welkos and I are joint inventors of the subject matter disclosed in the above-referenced application.
- 4. From [redacted date which is before 13 March 1996] to February 1998, I conducted research and development on a plague vaccine comprising a F1-V fusion protein as an immunogen as part of the Army Plague Vaccine Group.
- 5. Before about [redacted date which is before 13 March 1996], I obtained alhydrogel F1-V partial preparations from David G. Heath.
- 6. In my laboratory notebook I usually referred to the F1-V partial as "F1-V".
- 7. On [redacted date which is before 13 March 1996], I began mouse challenge studies with the F1-V partial preparations I obtained from David G. Heath. The experimental protocol for the challenge studies is provided in my notebook #3598 on page 123. See Exhibit GA1.
- 8. The results dated [redacted date which is before 13 March 1996] for the challenge studies using *Yersinia pestis* strain CO92, which is F1⁺ strain, with mice immunized alhydrogel F1-V partial preparations and controls with are provided in my notebook #3598 on pages 125-126. See Exhibit GA2.
- 9. The results dated [redacted date which is before 13 March 1996] for the challenge studies using *Yersinia pestis* strain C12, which is F1 strain, with mice immunized with alhydrogel F1-V partial preparations and controls with are provided in my notebook #3598 on pages 127-130. See Exhibit GA3.
- 10. On [redacted date which is before 13 March 1996], I wrote in my notebook #3598 on page 132, that the data on page 131 of my notebook and the mouse challenge studies are the first direct evidence that the F1-V fusion (F1-V partial) can induce an immune response to both F1 and V protein. David G. Heath witnessed this page and the results to which it references. See Exhibit GA4.
- 11. Exhibit AF3 (GA5) is an excerpt of my notebook #3598.
- 12. On [redacted date which is before 13 March 1996], I gave David G. Heath the protocol for formulating the F1-V whole vaccine preparations for mouse challenge assays. See Exhibit

DH16 (GA6).

- 13. In my laboratory notebook, I usually referred to the F1-V whole as "F1-WV".
- 14. Before about [redacted date which is before 13 March 1996], I obtained alhydrogel F1-V whole preparations from David G. Heath and began conducting the mouse challenge studies. See Exhibit GA7.
- 15. The results of the challenge studies dated [redacted date which is before 13 March 1996] using *Yersinia pestis* strain C12 or CO92 with mice immunized with alhydrogel F1-V partial preparations and controls are provided in my notebook #3739 on pages 60-63. The results show that F1-V whole confer a protective immune response against both F⁺ and F⁻ *Yersinia pestis* strains. See Exhibit GA8.
- 16. From about [redacted date which is before 13 March 1996] to 27 August 1996, I conducted further experiments to determine the efficacy of the F1-V fusion proteins and to determine whether any refinements could be made, such as the following:
 - a. On [redacted date which is before 13 March 1996], I conducted a mouse challenge study examining the long term efficacy of F1-V whole which is documented in my notebook #3739, page 75. See Exhibit GA9. The results dated [redacted date which is before 13 March 1996] are provided in my notebook #3739, page 85. See Exhibit GA10.
 - b. On [redacted date which is before 13 March 1996], I conducted a mouse challenge study examining the range of Al concentration which maintains an adequate adjuvant response which is documented in my notebook #3739, page 88. See Exhibit GA11. The alhydrogel F1-V whole preparations were obtained from David G. Heath. I copied the notebook pages from David G. Heath's notebook and inserted in my notebook. See Exhibit GA12. The results dated [redacted date which is before 13 March 1996] are found in my notebook #3739, pages 104-107. See Exhibit GA13.
 - c. Exhibit GA14 shows experimental data from my notebook #3739 which evidence that from [redacted date which is before 13 March 1996] to 23 February 1996, I conducted various mouse challenge studies with F1-V whole.
 - d. On 3 April 1996, I obtained the results for ELISA assays of serum obtained from mice immunized with F1-V whole to determine if the sera still contained antibodies against F1 antigen and V antigen as evidenced in my notebook #3739, page 122. See Exhibit GA15.
 - e. On about 15 May 1996, I conducted a study with mice vaccinated with different amounts of F1-V whole protein. The mice were challenged subcutaneously and by aerosol with Y. pestis, C092 or C12. See Exhibit GA16.
 - f. On about 28 June 1996, I conducted a study with mice vaccinated with either F1-V whole or a mixture of F1 + V or Plague USP vaccine. The mice were challenged by aerosol with Y. pestis C092. See pages 134 and 137 of my notebook #3739, Exhibit GA17.
 - g. On about 5 July 1996, I conducted a study with mice vaccinated with either F1-V whole or F1 + V or Plague USP vaccine. The mice were challenged

subcutaneously and by aerosol with Y. pestis, C092 or C12. See pages 135-136 of my notebook #3739, Exhibit GA18.

- 17. For all the challenge studies referenced herein, I obtained most of the *Yersinia pestis* strains C12 and CO92 from Susan L. Welkos.
- 18. On 15 February 1996, I presented the work summarized in David G. Heath's Abstract 17. See Exhibit DH19 (GA19).
- 19. I left the Army Plague Vaccine Group on 26 February 1998 when I retired from the U.S. Army.
- 20. I have reviewed and analyzed the Titball patent and the three priority documents, UK 9505059, UK 9518946, and UK 9524825, and PCT/GB96/00571.
- 21. It is my opinion that prior to 13 March 1996, the filing date of PCT/GB96/00571, the inventors of the Titball patent had not conceived and/or reduced to practice a plague vaccine comprising <u>purified</u> F1 antigen fused to all or part of V antigen as nowhere do UK 9505059, UK 9518946, and UK 9524825 disclose <u>isolating</u> or <u>purifying</u> a protein comprising F1 antigen fused to all or part of V antigen from the host cell and other cellular components and/or administering a purified protein comprising F1 antigen fused to all or part of V antigen to a subject.
 - a. In fact, UK 9518946 is the first disclosure indicating a genetic vaccine or how a host organism may be transfected with DNA for F1 antigen and V antigen to result in a live vaccine, i.e. an attenuated host organism (such as Salmonella) which produces the antigen when administered to a subject.
 - b. As described in UK 9518946, the genetic vaccine or the live vaccine is administered to a subject such that the protein/antigen of interest is then produced in the subject.
 - c. UK 9518946 does not describe isolating the protein/antigen of interest from the host organism and purifying the protein/antigen of interest from other cellular components prior to administration to a subject.
 - d. The genetic vaccine or live vaccine described in UK 9518946 is not a <u>purified</u> protein comprising F1 antigen fused to all or part of V antigen which is isolated and purified from cells and other cellular components as claimed in the above-referenced application.
- 22. I have reviewed and analyzed the experiments and data of the Army Plague Vaccine Group and it is my opinion that the Army Plague Vaccine Group:
 - a. Conceived of a fusion protein comprising F1 antigen fused to part of V by at least [redacted date which is before 13 March 1996].
 - b. Conceived of a fusion protein comprising F1 antigen fused to all of V by at least [redacted date which is before 13 March 1996].
 - c. Conceived of and reduced to practice a <u>purified</u> fusion protein comprising F1 antigen fused to part of V by at least [redacted date which is before 13 March 1996].
 - d. Conceived of and reduced to practice a purified fusion protein comprising F1

antigen fused to all of V by at least [redacted date which is before 13 March 1996].

- e. Conceived of and reduced to practice a vaccine against plague comprising a <u>purified</u> fusion protein comprising F1 antigen fused to part of V by at least [redacted date which is before 13 March 1996].
- f. Conceived of and reduced to practice a vaccine against plague comprising a <u>purified</u> fusion protein comprising F1 antigen fused to all of V by at least [redacted date which is before 13 March 1996].
- 23. I declare that all statements made herein of my own knowledge are believed to be true, and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under §1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

George W. Anderson, Jr.

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Date: 14 March 2007

CURRICULUM VITAE

George W. Anderson, Jr.

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E-mail: gwanders@bechtel.com

PROFESSIONAL EXPERIENCE:

2004-Present Senior Principal Engineer. Scientific responsibilities for the

Operations and Sustainment of a Defense Threat Reduction sponsored multi-country epidemiological surveillance system and collaborative

biological research program in the former Soviet Union.

2003-2004 Principal Advisor. Midwest Research Institute. General scientific

guidance to the company and responsibilities for integrating the capabilities of various company divisions in projects. Continue participation in DTRA projects in the former Soviet Union

(Kazakhstan, Uzbekistan and Republic of Georgia).

Senior Program Manager. Department at Southern Research Institute was purchased by Midwest Research Institute. I continued program management of the work in the former Soviet Union until promoted to

Principal Advisor.

1998-2003 Director, Medical Countermeasures Department. Southern Research

Institute. Responsible for the management, direction, control and review of the departmental research and development programs. Established a Biological Safety Level-3 containment facility for

vaccine potency testing of Department of Defense (DoD)

Investigational New Product (IND) vaccines (VEE, WEE, EEE, Q fever, Tularemia) in Frederick, MD. Department provided the DoD with expertise in biological defense, biosecurity, biotechnology and

biosafety for DARPA and DTRA projects at former biological weapons facilities in the former Soviet Union (FSU). Some of the projects support Non-proliferation efforts. Projects in the FSU include on-site observations of laboratory work in BSL-2, -3 and BSL-4 containment laboratories. Worked as a consultant to the Department of the Army for the anthrax vaccine production facility in the United States. Conducted audits in BSL-3 laboratories. Immunized with most licensed and IND bio-defense vaccines. Biosafety consultant to U.S. laboratories. Technical spokes person on biosafety for dismantlement of a pilot production facility for biological warfare agents in the United States.

1998-1998

Principal Scientist, SRS Technologies. Perform and manage scientific and technical tasks requiring the assessment of chemical (CW) or biological warfare (BW) programs/capabilities of foreign countries for terrorist groups and the development of recommended measures to curb the proliferation of biological weapons and technologies.

1993-1998

Chief, Pathogenesis and Immunology Branch, U.S. Army Medical Research Institute of Infectious diseases (USAMRIID). Branch Chief responsible for directing research programs directed toward development and production of prophylactic and therapeutic modalities against bacterial diseases of potential biological warfare significance (e.g., anthrax, plague, Q fever, tularemia, glanders). Special project officer for multi-million dollar Good Laboratory Practices project which included facility upgrades, Responsible for the GLP BSL-3 laboratory setup, and project manager for multi-year preclinical project for a supplement to the anthrax vaccine license. Research associate on the clinical protocol for a supplement to the current anthrax vaccine license. Manage technical aspects of a contract for cGMP production of cell banks and recombinant Bacillus anthracis PA protein as diagnostic or vaccine component. Involved with nonproliferation activities with former Soviet weapons scientist. First US military officer invited into the BSL-3 containment laboratories at the State Research Center of Applied Microbiology; Obolensk, Russia.

1990-1993

Medical R&D Officer, Science and Technology Center-Europe, Frankfurt, Germany - Responsible for finding biotechnologies, products, and collaboration in Europe, the Middle East, Africa, and the former Soviet Union, which could shorten or negate the need for the R&D cycle in Defense Department laboratories. This work involved extensive travel to scientific conference and Institutes in these geographical areas and technical report writing.

Curriculum vitae, George W. Anderson, Jr., 1/26/2007

1987-1990 Research Immunologist, U.S. Army Medical Research Institute of

Infectious Diseases. Investigations included development of a congenic strain of rats, vaccine efficacy trials against an aerosol exposure, and pathogenesis studies on Rift Valley fever virus. These

studies were carried out in a Biosafety Level 3 laboratory.

1983-1987 Graduate student at the Johns Hopkins University, while on active

duty, US Army.

1977-1983 Research Scientist, U.S. Army Medical Research Institute of

Infectious Diseases (USAMRIID), Fort Detrick, Frederick, Maryland - Was responsible for developing and investigating genetically define

animal models for exotic viral and rickettsial diseases.

ACADEMIC

APPOINTMENT: Associate Professor, Center for Disaster Preparedness, School of

Medicine, University of Alabama

FIELD STUDIES:

Member of a team of two who conducted an epidemiological sero-

survey for phleboviral infections at the MRC Laboratory, The Gambia.

and at the Institute Pasteur, Senegal, West Africa.

MILITARY SERVICE:

1977-1998 Retirement rank: LTC, U.S. Regular Army Commission, MSC

Army Management Staff College, graduate Command and General Staff College, graduate

EDUCATION:

Ph.D., 1988 The Johns Hopkins University, Baltimore, Maryland, Viral

Immunology. Dissertation: Viral and Host Determinants of

Resistance to Rift Valley Fever in a Rat Model

M.S., 1977 Florida Institute of Technology, Melbourne, Florida, Biology.

B.S., 1975 Florida Institute of Technology, Melbourne, Florida, Biology.

TEACHING EXPERIENCE:

1996 Mentor, Department of Defense, Science & Engineering Apprentice

Program

1989-1990 Served as a thesis committee member for a master's level candidate at

Hood College, Frederick, Maryland. Supervised the candidate's

research.

1988-1989 Sponsored a Korean ophthalmologist for the 1988-1989 ROK/US

Scientist/Engineer Exchange Program in my laboratory to develop a

model to study Rift Valley fever ocular sequelae.

1975-1977 Teaching assistantship at Florida Institute of Technology with primary

teaching responsibilities for microbiology and biochemistry

laboratories.

MEMBERSHIP IN ACADEMIC AND PROFESSIONAL SOCIETIES:

Membership:

American Society for Microbiology

Sigma XI Scientific Research Society

HONORS AND AWARDS:

Who's Who Among Students in American Universities – 1974, 1975

Four-year Army ROTC Scholarship

Four-year U.S. Army Long Term Health, Education and Training

Program (Ph.D. scholarship to the Johns Hopkins University)

Blue Key National Honor Fraternity

Sigma XI Scientific Research Society

Scouting: Highest rank - Eagle, Highest honor - Order of the Arrow,

Vigil Member

U.S. Army Army Commendation Medal

Meritorious Service Award w/2 Oak leaf clusters

Legion of Merit

Distinguished Professional Achievement Award, Florida Institute of

Technology 2002

Letter of appreciation from Joint Program Office for Biological

Defense for assistance to BioPort Corporation in obtaining FDA approval for Biological License Agreement for anthrax

vaccine

PATENTS:

U.S. Patent Number 5,320,069, "Small Animal Restraint Device"

Patent Pending Recombinant F1-V Plague Vaccine, filing #08/699,716

18 Dec 96

CONTINUING EDUCATION:

The Regulatory Process and Good Clinical Practices, Technology Management Integration, Inc., 1994

Regulatory Issues in Biotechnology, Univ. MD, 1995 Good Manufacturing Processes for Bioprocesses, Univ. MD, 1996 Quality Control and Quality Assurance of Biotechnology Products, Univ. MD, 1996

Assay Validation, PDA, 1996

Intro.to GLPs and Auditing, International Center for Health & Environmental Education, 1996

Writing and Evaluating Standard Operating Procedures for the Regulatory Environment, International Quality Training, 1996

Intro. FDA Good Laboratory Practices & Documentation Principles, International Quality Training, 1996

Good Laboratory Practices Regulations for Study Directors, International Quality Training, 1996

Fundamentals & Concepts of Calibration & Metrology, PDA, 1996 Biotechnology GMP Facility Design, Construction and Validation, Univ. MD, 1997

Advances in Filtration and Bioseparation Technologies, Pall Ultrafine Filtration Company, Columbia, MD, 1997

Validation of Biotechnology Processes and Systems, Univ. MD, 1997 Fermentation Microbiology, American Type Culture Collection Workshop, Rockville, MD, 1997

Fundamentals of D, F and Z Values, PDA, 1997

Basic Principles in Preparation of Sterile Dosage Forms, PDA, 1997 Parenteral Packaging: Rubber, Glass, Plastic and Metal Seals, PDA, 1997

Regulatory Compliance Training, Southern Research Institute, 1998 ISO 9001/Quality system Introductory Training, Southern Research Institute, 1998

Introductory to Earned Value Seminar, Dynport, LLC Professional Development, 1998

Positive Pressure Pneumatic BSL-4 Suite Training, State Research Center of Virology and Biotechnology, "Vector", Russia 2000 Introduction To Aerosol Mechanics I & II, AAAR, 2000

USA-Russia Workshop on International Research Ethics; Institutional Review Boards and Laboratory Animal Welfare, 2002

Introduction to Laboratory Ventilation and Design, American Biological Safety Association, 2002

Plant Biosafety, American Biological Safety Association, 2002 Bechtel Safety Leadership Workshop, Bechtel National Inc, 2006

The Transport of diagnostic & Infectious Samples, American Biological Safety Association, 2006

Biohazard Risk Assessment, American Biological Safety Association, 2006

Certifications:

Transport of diagnostic & Infectious Substances by Air (per ICAO Technical Instructions & IATA DGR) valid until 15 Oct 2008

REVIEWER/CONSULTANT:

U.S. Army In-house laboratory independent Research (ILIR) proposals, 1993-1998

U.S. Army Broad Agency Announcement proposals, 1993-1998 Experts Contact for database to assist Biological Arms Control Treaty Office (BACTO), 1997-1998

Nonproliferation Programs IntraAgency Roundtable, 1997-1998 U.S.-Uzbek Collaborative Biotechnology Grants Program, 2000 Consultant for DoD at BioPort Corporation (anthrax vaccine production facility), 2000-2002

PROFESSIONAL PUBLICATIONS:

- 1. Anderson, G.W., Jr., and J.V. Osterman. 1980. Host defenses in experimental Rickettsialpox: Genetics of natural resistance to infection. Infect. Immun. 28: 132-136.
- 2. **Anderson, G.W., Jr.**, and J.V. Osterman. 1980. Host defenses in Rickettsialpox: Resistance of C3H mouse sublines. Acta. Virol. 24: 294-296.
- 3. Peters, C.J., and **G.W. Anderson, Jr.** 1981. Pathogenesis of Rift Valley fever, pp. 21-41. In Contributions to Epidemiology and Statistics, vol. 3. (N. Goldblum, T.A. Swartz, and M.A. Kingberg, eds.). S. Karger, Basel.
- 4. Anderson, G.W., Jr., T.W. Slone, Jr., and C.J. Peters. 1987. Pathogenesis of Rift Valley fever virus (RVFV) in inbred rats. Microb. Pathogen. 2: 283-293.
- 5. **Anderson, G.W., Jr.**, and J.F. Smith 1987. Immunoelectron microscopy of Rift Valley fever viral morphogenesis in primary rat hepatocytes. Virology. 161: 91-100.
- 6. Anderson, G.W., Jr., and C.J. Peters. 1988. Viral determinants of virulence for Rift Valley fever (RVF) in rats. Microb. Pathogen. 5: 241-250.
- 7. **Anderson, G.W., Jr.,** T.W. Slone, Jr., and C.J. Peters. 1988. The gerbil, *Meriones unguiculatus*. A model for Rift Valley fever viral encephalitis. Archives Virology 102: 187-196.
- 8. Anderson, G.W., Jr., J.-F. Saluzzo, T.G. Ksiazek, J.F. Smith, W. Ennis, D. Thureen, C.J. Peters, and J.P. Digoutte. 1989. Comparison of in vitro and in vivo systems for propagation of Rift Valley fever virus from clinical specimens. Res. Virol. 140:129-138.
- 9. Saluzzo, J.F., **G.W. Anderson, Jr.**, L.A. Hodgson, J.P. Digoutte, and J.F. Smith. 1989. Antigenic and biological properties of Rift Valley fever virus isolated during the 1987 Mauritanian epidemic. Res. Virol. 140:155-164.
- 10. Peters, C.J., C.-T. Liu, **G.W. Anderson, Jr.**, J.C. Morrill, and P.B. Jahrling. 1989. Pathogenesis of viral hemorrhagic fevers: Rift Valley fever and Lassa fever contrasted. Rev. Infect. Dis. 11 (Suppl. 4): 5743-5749.
- 11. Saluzzo, J.F., G.W. Anderson, Jr., J.F. Smith, D. Fontenille, and P. Coulanges. 1989. Biological and antigenic relationship between Rift Valley fever virus strains isolated in Egypt and Madagascar. Trans. R. Soc. Trop. Med. Hyg. 83:701.
- 12. Solow, R., K. Mereish, G.W. Anderson, Jr., and J. Hewetson. 1990, Effect of microcystin-LR on cultured rat endothelial cells. Med. Sci. Res. 18:241-244.

- 13. Anderson, G.W., Jr., M.V. Slayter, W. Hall, and C.J. Peters. 1990. Pathogenesis of a phleboviral infection (Punta Toro virus) in Golden Syrian hamsters. Arch. Virol. 114: 203-212.
- Anderson, G.W., Jr., J.O. Lee, A.O. Anderson, N. Powell, J.A. Mangiafico, and G. Meadors. 1991. Efficacy of a Rift Valley fever virus vaccine against an aerosol infection in rats. Vaccine. 9: 710-714.
- 15. **Anderson, G.W., Jr.**, J.A. Rosebrock, A.J. Johnson, G.B. Jennings, and C.J. Peters. 1991. Infection of inbred rats with Rift Valley Fever virus: Development of a congenic resistant strain and observations on age-dependence of resistance. Am. J. Trop. Med. Hyg. 44(5): 475-480.
- 16. Anderson, G.W., Jr., W.B. Lawrence, J-O Lee, and M. Young. 1991. A restraint for ophthalmic examination of unanesthetized rats. Note. Laboratory Animal Science. 41(3): 288-290.
- 17. Friedlander, A.M., S.L. Welkos, P.L. Worsham, G.P. Andrews, D.G. Heath, G.W. Anderson, Jr., M.L.M. Pitt, J. Estep, and K. Davis. 1995. Relationship between virulence and Immunity as revealed in recent studies of the F1 capsule of Yersinia pestis. Clinical Infectious Diseases. 21(Suppl 2):S178-81.
- 18. Andrews, G.P., D.G. Heath, **G.W. Anderson**, **Jr.**, S.L. Welkos, and A.M. Friedlander. 1996. Fraction 1 capsular antigen (F1) purification from *Yersinia pestis* CO92 and an *Escherichia coli* recombinant strain and efficacy against lethal plague challenge. Infect. Immun. 64:2180-2187.
- 19. Anderson, G.W. Jr., S.E.C. Leary, E.D. Williamson, R.W. Titball, S.L. Welkos, P.L. Worsham, and A.M. Friedlander. 1996. Recombinant V antigen protects mice against pneumonic and bubonic plague caused by F1-capsule-positive and -negative strains of *Yersinia pestis*. Infect. Immun. 64:4580-4585.
- 20. Anderson, G.W. Jr., P.L. Worsham, C.R. Bolt, G.P. Andrews, S.L. Welkos, A.M. Friedlander, and J.P. Burans. 1997. Protection of mice from fatal bubonic and pneumonic plague by passive immunization with monoclonal antibodies against the F1 protein of *Yersinia pestis*. Am. J. Trop. Med. Hyg. 64:4580-4585.
- 21. Heath, D.G., **G.W. Anderson, Jr.**, S.L. Welkos, A.M. Friedlander, and J.M. Mauro. 1997. A recombinant capsular F1-V antigen fusion protein vaccine protects against experimental bubonic and pneumonic plague. *in* Vaccines 97. Cold Spring Harbor Laboratory Press, pp 197-200.
- 22. Pullen, J.K., **G.W. Anderson, Jr.**, S.L. Welkos, and A.M. Friedlander. 1998. Analysis of the *Yersinia pestis* V protein for the presence of linear antibody epitopes. Infect. Immun. 66:521-527.

- 23. Anderson, G.W. Jr., D.G. Heath, C.R. Bolt, S.L. Welkos, and A.M. Friedlander. Short-and long-term efficacy of single-dose subunit vaccines against *Yersinia pestis* in mice. Am. J. Trop. Med. Hyg., 58(6): 793-799.
- 24. Ivins, B.E., M.L.M. Pitt, P.F. Fellows, J.W. Farchaus, G.E. Benner, D.M. Waag, S.F. Little, **G.W. Anderson, Jr.**, P.H. Gibbs, and A.M. Friedlander. 1998. Comparative efficacy of experimental anthrax vaccine candidates against inhalation anthrax in rhesus macaques. Vaccine, 16(11/12): 1141-1148.
- 25. Heath, D.G., **G.W. Anderson, Jr.**, J.M. Mauro, S.L. Welkos, G.P. Andrews, J. Adamovicz, and A.M. Friedlander. 1998. Protection against experimental bubonic and pneumonic plague by a recombinant capsular F1-V antigen fusion protein vaccine. Vaccine, 16(11/12): 1131-1137.
- 26. Andrews, G.P., S.T. Strachan, G.E. Benner, A.K. Sample, J.J. Adamovicz, G.W. Anderson, Jr., S.L. Welkos, A.M. Friedlander. 1999. Protective efficacy of recombinant *Yersinia* outer proteins (Yops) against bubonic plague caused by encapsulated and non-encapsulated *Yersinia pestis*. Infect. Immun., 67(3): 1533-1537.
- 27. Lawrence W.B., **G.W. Anderson, Jr.**, J.0. Lee, and W.C. Hall. Ocular sequelae associated with Rift Valley fever virus (RVFV) infection in inbred rats (submitted).

PUBLISHED ABSTRACTS:

- 1. Rosebrock, J.A., G.W. Anderson, Jr., H. Schellekens, and C.J. Peters. 1983. Differential interferon sensitivities of lethal and non-lethal strains of Rift Valley fever virus (RVFV) in vitro. In vitro 19: 286-287.
- 2. Anderson, G.W., Jr., M.V. Slayter, and C.J. Peters. 1988. Pathogenesis of a phleboviral infection (Punto Toro virus) in golden Syrian hamsters. Virus Supplement 2: 40.
- 3. Ribas, J.L., M.D. Kanzer, G.W. Anderson, Jr., J. Sesterhenn, and C.J. Peters. 1989. Rift Valley fever viral encephalitis in the gerbil. J. Neuropathol. Exp. Neurol. 48: 315.
- 4. Ribas, J.L., M.D. Kanzer, G.W. Anderson, Jr., E. Perez-Rosario, and C.J. Peters. 1990. Rift Valley fever viral encephalitis in the gerbil: Ultrastructural and immunocytochemical correlation. J. Neuropath. Exp. Neurol. 49:348(A).

Other publications:

Technical Report for Alternate Air Collection Media, SRS Technologies, TR98-156

PRESENTATIONS:

- 1. Anderson, G.W., Jr., and J.V. Osterman. Susceptibility of mouse strains to *Rickettsia akari*. Presented at the 79th Annual Meeting of the American Society of Microbiologists, Los Angeles, California, 1979.
- 2. Anderson, G.W., Jr., and J.V. Osterman. Susceptibility of mice to *Rickettsia akari*. Presented at the 1st Annual Rickettsiology Conference. Port Deposit, Maryland, 1979.
- 3. Anderson, G.W., Jr., and C.J. Peters. Effect of immunosuppression on genetically resistant LEWIS/Mai rats to Rift Valley fever virus. Presented at the Maryland-D.C. Branch of the American Society for Microbiology Meeting, Fort Detrick, Frederick, Maryland, January 1981.
- 4. Anderson, G.W., Jr., C.J. Peters, and T.W. Slone. Pathogenesis of Rift Valley fever virus in inbred rats. Presented at the 81st Annual Meeting of the American Society for Microbiology, Dallas, Texas, March 1981. Abstracts of the Meeting, D244.
- 5. Peters, C.J., and **G.W. Anderson**, **Jr.** Pathogenesis of Rift Valley fever and other Phlebovirus infections. Presented at the 5th International Congress of Virology, Strasbourg, France, August 1981.
- 6. Peters, C.J., and **G.W. Anderson**, **Jr.** Resistance to Phleboviruses. Presented at the U.S.-Japan Cooperative Medical Science Program, Bethesda, Maryland, 9 November 1981.
- 7. Peters, C.J., and **G.W. Anderson, Jr.** Pathogenesis of Phlebovirus infections. Presented at the 30th Annual Meeting of the American Society of Tropical Medicine and Hygiene, San Juan, Puerto Rico, November 1981.
- 8. Anderson, G.W., Jr., and C.J. Peters. Role of humoral immunity in Rift Valley fever infection. Presented at the 49th Conjoint Meeting on Infectious Diseases, Ontario, Canada, 25 November 1981.
- 9. Anderson, G.W., Jr., J.A. Rosebrock, A.J. Johnson, and C.J. Peters. Age- and dose-dependent resistance of rats to Rift Valley fever virus. Presented at the 82nd Annual Meeting of the American Society for Microbiology, Atlanta, Georgia, March 1982.
- 10. Anderson, G.W., Jr., T,W. Slone, Jr., and C.J. Peters. A model for the encephalitic form of Rift Valley fever. Presented at the 31st Annual Meeting of the American Society of Tropical Medicine and Hygiene, Cleveland, Ohio, 1982.
- 11. Peters, C.J., H. Schellekens, J.A. Rosebrock, and G.W. Anderson, Jr. Genes, macrophages, and resistance to Rift Valley fever in the rat. Presented at the First Annual Meeting of the American Society for Virology, Ithaca, New York, 1982.

- 12. Peters, C.J., H. Schellenkens, J.A. Rosebrock, and **G.W. Anderson, Jr.** Genetic resistance to Rift Valley fever virus: Role of macrophages and interferon. Fourth International Conference on Comparative Virology, Banff, Canada, October 1982.
- 13. Anderson, G.W., Jr., and J.F. Smith. Rift Valley fever virus (RVFV) maturation at the plasma membrane of rat hepatocytes as revealed by immunoelectron microscopy. Presented at the 35th Annual Meeting of the American Society of Tropical Medicine and Hygiene, Denver, Colorado, 8-11 December 1986.
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- 37. Anderson, Jr. G.W., Laboratory/Field Biosafety. Presented at the Veterinary and Human Brucellosis Workshop, Almaty, Kazakhstan, 19-22 July 2004.
- 38. Anderson, Jr. G.W., Collaborative Biosafety Efforts between the Defense Threat Reduction Agency (DTRA) and Russian Institutes. Presented at the Development of International Collaboration in Infectious Disease Research Conference, Novosibirsk, Russia, 8-10 September 2004.

File: F1-V fus Protocol: B9	sion last update REDACTED 4-02 rotein immunization and challenge	i	
	rotein immunization and challenge		
•	Jy		
Investigators:	CPT Heath, Dr. Welkos, LTC Anderson, COL Friedlander		
is positive by	CPT David Heath has produced and purified a recombinant F1-V Western blot to F1 and V. Antigen dose will be based on 10 μg I to it. Subcutaneous injection at nape of neck.	/ fusion pr F1/dose -	otein. The protein + the amount of V
V-antigen use Heath's noteb	d in this experiment is from Mauro. Details of the V-antigen ca	n be obtai	ined from CPT
Purpose: Imm strain of Y. po	nunize and challenge mice to check on immunogenicity and prote estis by sc and aerosol challenge, 50 LD ₅₀ .	ection agai	nst the CO92
EcF1c will be	endotoxin free, same F1 as used in the active immunization e	xperiment	
Alhydrogel, 1.	3%, from SuperFos. Batch <u># 2043</u> , Expiration date <u>None</u> ,	μg	of AL/dose
Immunization	Groups: 10 Swiss Webster female mice per group from Harlan	Sprague [Dawley
	he amount of F1 will be held constant.		
Subcutaneous		Strain	# Mice
	Alhydrogel alone, days 0, 30, sc	CO92	10
Group 2	alhydrogel + 10 µg F1, days 0, 30, sc	CO92	10
	alhydrogel + 10 μg F1 urea treated, days 0, 30, sc	CO92	10
Group 4	Alhydrogel + 18.5 μg F1-V fusion protein days 0, 30, sc	CO92	10
Aerosol challe	nge		
Group 5	Alhydrogel alone, days 0, 30, sc	CO92	10
Group 6	alhydrogel + 10 μg F1, days 0, 30, sc	CO92	10
Group 7	alhydrogel + 10 μg F1 urea treated, days 0, 30, sc	CO92	10
Group 8	Alhydrogel + 18.5 μg F1-V fusion protein, days 0, 30, sc	CO92	10
Group 9	Alhydrogel + 37.0 μg F1-V fusion protein days 0, 30, sc	CO92	<u>10</u>
		Total	90
1F622A6174	/F-V-000		
1F633B5D66			
1F666A563B			
1F684B1B13			•
1F73143D1D		•	
1F6336192F/			
1F617A0402	F-V-006		
1F6373612A			
1F642A0E45/			
1F64667324/	<u>F-V-009</u>		
1F610F757C/	F-V-010		
1F625B356F/	F-V-011		

1F62584463/F-V-012 2007432B6B/F-V-013 1F620D6B07/F-V-014 1F65793A49/F-V-015

REDACTED

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1F61041D5F/F-V-016
  1F59570130/F-V-017
  1F62776325/F-V-018
  1F6E2E3015/F-V-019
  1F60055D1F/F-V-020
  1F66337F49/F-V-021
  1F6E25735B/F-V-022
  1F647B4141/F-V-023
  1F6835251F/F-V-024
 1F767F7A72/F-V-025, Not responding replaced with 1F617C1371
 1F62530428/F-V-026
 1F63717815/F-V-027
 1F65122842/F-V-028
 1F666F1F6D/F-V-029
 1F56376371/F-V-030
 1F66655A3C/F-V-031
 1F663A6859/F-V-032
 1F64163037/F-V-033
 1F72107B64/F-V-034
 1F63467B3D/F-V-035
 1F645B1111/F-V-036
 1F61524668/F-V-037
 1F6132321C/F-V-038
 1F655A0E14/F-V-039
 1F661E3726/F-V-040, Not responding, replaced with 1F6134004C
 1F656C5937/F-V-041
 1F61135D10/F-V-042
 1F6655574F/F-V-043
 1F664B210F/F-V-044
 7F7B0A2C5B/F-V-045
 1F63737516/F-V-046
 1F66240C4B/F-V-047
1F5A6F0C0C/F-V-048
1F65354106/F-V-049
1F76011A50/F-V-050
1F653D7946/F-V-051
1F624D6A48/F-V-052
1F64030B6F/F-V-053
1F6E295D6D/F-V-054
1F64303B12/F-V-055
1F635D326F/F-V-056
1F6726084C/F-V-057
1F6317796E/F-V-058
1F64024437/F-V-059
1F66056C0A/F-V-060, died from anesthesia on17FEB95 during bleeding
1F637E6917/F-V-061
1F64653F59/F-V-062
1F643F7846/F-V-063
1F6412204B/F-V-064
7F7D261230/F-V-065
1F6132735B/F-V-066
1F635C5D45/F-V-067
1F636A1103/F-V-068
1F5F542F7F/F-V-069
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1F6729527F/F-V-070 1F63280353/F-V-071 1F600B6D09/F-V-072 1F77037275/F-V-073 1F615E071B/F-V-074 1F646F0D01/F-V-075 1F66415A60/F-V-076 1F65566046/F-V-077 1F64495262/F-V-078 1F6467682E/F-V-079 200905054D/F-V-080 1F636F2F60/F-V-081 1F650E541A/F-V-082 1F637B3E45/F-V-083 1F673C5668/F-V-084 1F647E2E51/F-V-085 1F663B0937/F-V-086 1F62111658/F-V-087 1F6320124C/F-V-088 1F635A1F05/F-V-089

CFA from Sigma Cat#F-5881 Lot#80H8808, 10 ml/bottle IFA from Sigma Cat#F-5506 Lot#80H8812, 10 ml/bottle

Groups 10-23 will be started one week later. The amount of V will be held constant, though the V part of the fusion protein is only about half the size of the native V.

Subcutaneou	s challenge	Strain	# Mice
Group 10	Alhydrogel alone, days 0, 30, sc	C12	10
Group 11	Alhydrogel days + 27 μg F1-V fusion protein day 0, 30, sc	C12	10
Group 12	CFA + 10 μg V, days 0, IFA 30, ip	C12	10
Group 13	CFA + 10 μg V urea treated, days 0, IFA 30, ip	C12	10
Group 14	CFA + 27 μg F1-V fusion protein days 0, IFA 30, ip	C12	10
Group 15	CFA + 54 μg F1-V fusion protein days 0, IFA 30, ip	C12	10
Group 16	CFA day 0, IFA day 30 alone, ip	C12	10
Aerosol chall	enge		
Group 17	Alhydrogel alone, days 0, 30, sc	C12	10
Group 18	Alhydrogel + 27 μg F1-V fusion protein, days 0, 30, sc	C12	10
Group 19	CFA + 10 μg V, days 0, IFA 30, ip	C12	10
Group 20	CFA + 10 μg V urea treated, days 0, IFA30, ip	C12	10
Group 21	CFA + 27 μg F1-V fusion protein day 0, IFA 30, ip	C12	10
Group 22	CFA + 54 μg F1-V fusion protein day 0, IFA 30, ip	C12	10
Group 23	CFA day 0, IFA day 30 alone, ip	C12	<u>10</u>
_		Total	140
Group 24	CEA + 27 up E1-V fusion protoin day 0 IEA 20 in contibody		40

Group 24 CFA + 27 μg F1-V fusion protein day 0, IFA 30, ip --antibody response 10 Measure titer at 14, 27,57

Chip # for Groups 10-24, Group 10 has some mice doubled 1F646A0D06/F-V-090 7F7B107C58/F-V-091 or 1F63125319, DOUBLE CHIPPED 1F6E345D62/F-V-092 OR 1F64791E66, DOUBLE CHIPPED

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1F64141752/F-V-093 OR 1F61116E01, DOUBLE CHIPPED
1F65020D6D/F-V-094 OR 9F7D25797E, DOUBLE CHIPPED
1F62032B51/F-V-095 OR 1F650627AF, DOUBLE CHIPPED
1F635D4B56/F-V-096 OR 7F7D243700, DOUBLE CHIPPED
7F7B06623C/F-V-097 OR 7F7D17224D, DOUBLE CHIPPED
7F7D23252D/F-V-098
1F630A7004/F-V-099
1F6458061F/F1-VB-001
1F66294012/F1-VB-002
1F65323119/F1-VB-003
1F68597E22/F1-VB-004
1F663B023E/F1-VB-005
1F64742F5A/F1-VB-006
1F62713757/F1-VB-007
1F664A1B16/F1-VB-008
1F624C3D76/F1-VB-009
1F60466754/F1-VB-010
1F650D343B/F1-VB-011
1F640F0668/F1-VB-012
1F684A6D42/F1-VB-013
1F627B4440/F1-VB-014
1F66362421/F1-VB-015
1F64121C4F/F1-VB-016
1F647A4340/F1-VB-017
1F757C7977/F1-VB-018
1F65625644/F1-VB-019
1F65662E68/F1-VB-020
1F655A455D/F1-VB-021, found dead, 13JAN95, cause unknown
1F63624B51/F1-VB-022
1F65487440/F1-VB-023
1F61185F09/F1-VB-024
1F63296273/F1-VB-025
1F6626094C/F1-VB-026
1F622A1C39/F1-VB-027
1F6122312D/F1-VB-028
1F6329775E/F1-VB-029
1F64652276/F1-VB-030
1F73061751/F1-VB-031
1F63672C6B/F1-VB-032
1F68406158/F1-VB-033
1F6359061F/F1-VB-034
1F653E2C12/F1-VB-035
1F60524B64/F1-VB-036
1F67274A09/F1-VB-037
200F13231B/F1-VB-038
1F620B7B79/F1-VB-039
1F66121653/F1-VB-040
1F66247661/F1-VB-041
1F652F0D40/F1-VB-042
1F63103B33/F1-VB-043
1F673D0C31/F1-VB-044
1F62757218/F1-VB-045
1F63401727/F1-VB-046
1F665F5D3F/F1-VB-047
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1F66585F44/F1-VB-048 1F61195611/F1-VB-049 200737366C/F1-VB-050 1F614E2012/F1-VB-051 1F64297C58/F1-VB-052 1F75463175/F1-VB-053 1F626E157C/F1-VB-054 1F650F5419/F1-VB-055 1F64075C1A/F1-VB-056 1F756A3151/F1-VB-057 1F6121124D/F1-VB-058 1F655E405E/F1-VB-059 1F65233C1D/F1-VB-060 1F683C0B32/F1-VB-061 1F615F021F/F1-VB-062 1F71772257/F1-VB-063 1F644F713D/F1-VB-064 1F67412415/F1-VB-065 1F66606C2F/F1-VB-066 1F77087969/F1-VB-067 1F637D1F62/F1-VB-068 1F61172742/F1-VB-069 1F64511E0E/F1-VB-070 1F5756496B/F1-VB-071 1F681C4617/F1-VB-072 1F68571012/F1-VB-073 1F682F6763/F1-VB-074 1F673E4577/F1-VB-075 1F62681205/F1-VB-076 1F655F1E7F/F1-VB-077 1F65523377/F1-VB-078 1F61284810/F1-VB-079 1F6152307E/F1-VB-080 1F68382A17/F1-VB-081 1F686B1876/F1-VB-082 1F60106E03/F1-VB-083 1F63242D2D/F1-VB-084 1F637A7311/F1-VB-085 1F631F0D52/F1-VB-086 1F6378473F/F1-VB-087 1F63390C39/F1-VB-088 1F66517931/F1-VB-089 1F642C272A/F1-VB-090 1F654E703E/F1-VB-091 1F64134129/F1-VB-092 1F63355772/F1-VB-093 1F621A0263/F1-VB-094 1F64361A2D/F1-VB-095 1F61774A3F/F1-VB-096 1F65276075/F1-VB-097 1F646F0905/F1-VB-098 1F624C181B/F1-VB-099 1F684E3C6F/F1-VB-100

1F64034B2F/F1-VB-101

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1F65083C38/F1-VB-102
1F65792E55/F1-VB-103
1F6012412E/F1-VB-104
1F68321F28/F1-VB-105
1F6E313909/F1-VB-106
1F614D6F44/F1-VB-107
1F62394402/F1-VB-108
1F627B5C28/F1-VB-109
1F64165A0D/F1-VB-110
1F630B4231/F1-VB-111
1F6629450D/F1-VB-112
1,F5F630D12/F1-VB-113
1F657C5B25/F1-VB-114
1F6337596E/F1-VB-115
1F66297260/F1-VB-116
1F66524F5A/F1-VB-117
1F6829537D/F1-VB-118
1F64616E2E/F1-VB-119
1F61517936/F1-VB-120
1F65136504/F1-VB-121
1F66002A51/F1-VB-122
1F5F7D1075/F1-VB-123
1F65074134/F1-VB-124
1F71724836/F1-VB-125
1F656C4F41/F1-VB-126
1F60182148/F1-VB-127
1F6866573C/F1-VB-128
1F62780205/F1-VB-129
1F65080173/F1-VB-130
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Schedule
Groups 1-9
22NOV94
              Arrival of Swiss Webster mice, female 7-8 wks, Harlan Sprague Dawley in
                AA-3 (Barrier)
01DEC94
              Chipped with BioMedic Data Systems transponders in AA-3
16DEC94
              1st immunization, AA-3
13JAN95
              2nd Immunization, AA-3, day 28
17FEB95
              Bleed to determine prechallenge titers, AA-3, day 63, serum#1500-1589
24FEB95
              Challenge by aerosol & sc routes, day 70
              Terminal bleed, day 28 pi, titrate spleens, serum # 24
24MAR95
                                                      Tum # 9378 - 9432
Groups 10-23
29NOV94
              Arrival of Swiss Webster mice, female 7-8 wks, Harland Sprague Dawley in AA-3
              (Barrier)
13DEC94
              Chipped with BioMedic Data Systems transponders in AA-3 (SGT Zimmerman, Vet Med)
              1st immunization, AA-3; Crow & Fritzgerald helped
22DEC94
              2nd immunization, AA-3, day 29
20JAN95
              Bleed to dermine prechallenge titers, AA-3, day 64, serum# /628 - /7
24FEB95
              Challenge by aerosol & sc route, day 71
3MAR95
              Terminal bleed, day 28 pi, titrate spleens, serum # 1759 - 1800
31MAR95
                                                     Trace # 9433 - 9479
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Group 24

22DEC94

1st immunization, AA-3

06JAN95 20JAN95 Bleed, day 14, AA-3, SERUM# 1220-1229 Bleed, day 27, AA-3, SERUM# 1339-1348

20JAN95

2nd immunization, day 28, AA-3

24FEB95

Bleed, day 63, AA-3, SERUM# /76F- 1778 and bronchial lavage #

Exhibit GA2

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REDACTED Project: Active Immunization F1-V Alhydorgel lpocitium: Yersinia pestis strain CO92 RUN 2 - 104 LOSO erôso Dose: RUN 1 80 4000 Swiss Webster at 7-8wk Vendor: Harlan Sprague Dawley REDACTED REDACTED ay 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 24 25 26 27 28 Day postinfection 3 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 Comments/Chip # Group Cage# Alh alone 1F6134004C/F-V-040 GP5 Ĺ 1F656C5937/F-V-041 1F61135D10/F-V-042 1F6655574F/F-V-043 1F664B210F/F-V-044 7F7B0A2C5B/F-V-045 1F63737516/F-V-046 1F66240C4B/F-V-047 1F5A6F0C0C/F-V-048 1F65354106/F-V-049 10 Alh+F1 1F76011A50/F-V-050 10 µg 12 1F653D7946/F-V-051 GP6 13 1F624D6A48/F-V-052 1F64030B6F/F-V-053 RUK 1F6E295D6D/F-V-054 1F64303B12/F-V-055 1F635D326F/F-V-056 1F6726084C/F-V-057 1F6317796E/F-V-058 20 1F64024437/F-V-059 Alh+F1 Died during bleeding of 17FEB95 4F66056C0A/F-V-060 10 µg 22 1F637E6917/F-V-061 urea 1F64653F59/F-V-062 GP7 . 1F643F7846/F-V-063 25 100 1F6412204B/F-V-064 26 7F7D261230/F-V-065 27 1F6132735B/F-V-066 1F635C5D45/F-V-067 1F636A1103/F-V-068 1F5F542F7F/F-V-069 30 Alh+F1-V 1F6729527F/F-V-070 18.5 µg 1F63280353/F-V-071 GP8 1F600B6D09/F-V-072 1F77037275/F-V-073 1F615E071B/F-V-074 1F646F0D01/F-V-075 37 1F66415A60/F-V-076 1F65566046/F-V-077 39 1F64495262/F-V-078 1F6467682E/F-V-079 Alb+F1-V 200905054D/F-V-080 37 дд 1F636F2F60/F-V-081 GP9 1 1F650E541A/F-V-082 1F637B3E45/F-V-083 1F673C5668/F-V-084 1F647E2E51/F-V-085 1F663B0937/F-V-086 48 T 1F62111658/F-V-087 1 1F6320124C/F-V-088 1 1F635A1F05/F-V-089 For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions Discard dead animals Use scanner to check chip # of dead animals Mark number of mice alive Attno LTC Anderson

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Project: Active Immunization F1-V Alhydrogel/CFA DOOK #: 3598 Hum: Yersinia pestis strain C12 Dose: el enfosol Age: Arrive REDACTED Vendor: Harlan Sprague Dawley wiss Webster Sex: female DACTED by . 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 3 4 5 6 7 nostinfection. 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 Comments/Chip # 2 3 7 Cege# 51 1F683C0B32/F1-VB-061 52 / 1F615F021F/F1-VB-062 53 1F71772257/F1-VB-063 54 1F644F713D/F1-VB-064 55 1F67412415/F1-VB-065 1F66606C2F/F1-VB-066 1F77087969/F1-VB-067 58 1F637D1F62/F1-VB-068 59 1F61172742/F1-VB-069 60 1F64511E0E/F1-VB-070 61 1F5756496B/F1-VB-071 **7**62 1F681C4617/F1-VB-072 1F68571012/F1-VB-073 18 64 1F682F6763/F1-VB-074 1F673E4577/F1-VB-075 110 1F62681205/F1-VB-076 10 1F655F1E7F/F1-VB-077 1F65523377/F1-VB-078 69 1F61284810/F1-VB-079 1F6152307E/F1-VB-080 1F68382A17/F1-VB-081 1F686B1876/F1-VB-082 1F60106E03/F1-VB-083 1F63242D2D/F1-VB-084 1F637A7311/F1-VB-085 ┰ 1F631F0D52/F1-VB-086 1F6378473F/F1-VB-087 1F63390C39/F1-VB-088 79 11 1F66517931/F1-VB-089 1F642C272A/F1-VB-090 1F654E703E/F1-VB-091 1F64134129/F1-VB-092 83 1F63355772/F1-VB-093 1F621A0263/F1-VB-094 1F64361A2D/F1-VB-095 1F61774A3F/F1-VB-096 1F65276075/F1-VB-097 88 1F646F0905/F1-VB-098 1F624C181B/F1-VB-099 A 90 6 1F684E3C6F/F1-VB-100 91 1F64034B2F/F1-VB-101 92 1F65083C38/F1-VB-102 93 1F65792E55/F1-VB-103 1F6012412E/F1-VB-104 95 1F68321F28/F1-VB-105 96 1F6E313909/F1-VB-106 97 ſ 1F614D6F44/F1-VB-107 98 1F62394402/F1-VB-108 99 1F627B5C28/F1-VB-109 100 1F64165A0D/F1-VB-110 =1-V 101 1F630B4231/F1-VB-111 1 D 102 1F6629450D/F1-VB-112 103 1F5F630D12/F1-VB-113 104 1 1 1F657C5B25/F1-VB-114 105 DIE e 1F6337596E/F1-VB-115 106 107 108 1F66297260/F1-VB-116 1F66524F5A/F1-VB-117 1F6829537D/F1-VB-118 109 1F64616E2E/F1-VB-119 #110) 1 1 1F61517936/F1-VB-120 nimal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions d dead animals anner to check chip # of dead animals rumber of mice alive

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For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions Discard dead animals

Use scanner to check chip # of dead animals

Mark number of mice alive

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Notebook #: 3598 Incolum: Yeming pestis strain C12 REDACTED at 7-6wk Vendor: Harten Sprague Dawley Sex: female Sex:	Date RED	ACTED	Pro	ect	A	ctive	Imr	mun	izati	on F	1-7	Alh	ydro	gel/C	CFA																		
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For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions			竹	1	Ħ		1	1	1	17	17	17	T	T	lì	I	Li	í	ï	1		1	Li.	\prod			1			I i	1		200737366C/F1-VB-050
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Discard dead animals

Use scanner to check chip # of dead animals

Mark number of mice alive

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DREDA		Pro	ect:	Ac	ctive	lm	mun	izati	on F	1-V	Alh	ydro	gel/	CFA										_					_		_	
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For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions Discard dead animals

Use scanner to check chip # of dead animals

Mark number of mice alive

ELISA, FI & VANT.

STARTING DILUTION 1:640, SERIAL 1:2 DILUTIONS

-				GRALS
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FL PLATE "	SURUM#	VPATE#	FI PLATE #	JERUM# V	PLATE #
IAF	1117		IIA	1844	264
B	1118	\$ 16B		1845	В
2.A	1119	17A	12A	PET 1846	27.4
В	1230	3539 17B	.	1847	В
<i>3A</i>	1231	≥ 18A	13A	脚 1848	28/4
В	1232	₹118B	B	1849	B
4A	1233	19A	14 A	F# 1850	294
В	12.34	198	В	E NEG(N.	m.)
5A	1235	204	15A	F1+	304
<u> </u>	-1236- 18	36(FI) 20B	3 B	83_/	В
6A	-1237 18	37 (F1) 21A			
B	1238 18	38 (FI) 21B	PLATE 1-3 (F	i):	
7A	1836	1236 (FI) 22A	ا 10 المر200	0	
	1837 1	237(FI) 228	100 110 1	10	
8A	1838 1	238 (FI) 23A	REMOVED BUFFE	R FROM COLUMN	15 2-12
<i>B</i>	18	39 23B	ADDED 100pl KACI	H SAMPLE TO COLUM	NA 100 PUFFER
9A	摩 18	40 24A	TO COLUMNS 2	-12 , & RETITI	RATED SAMPLES
<i>B</i>	18	41 248		• :	
10A	18	42 25A			
B	185 189	43 258		· · · · · · · · · · · · · · · · · · ·	

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1836	POOL GP10 Alhydrogel alone	REDAC
1837	POOL GP11 Alh+27ugF1-V	T-0
1838	POOL GP12 CFA+10ugV	TED
1839	POOL GP13 CFA+10ugV,urea	1
1840	POOL GP14 CFA+27ugF1-V	1
1841	POOL GP15 CFA+54ugF1-V]
1842	POOL GP16 CFA alone	
1843	POOL GP17 Alhydrogel alone	
1844	POOL GP18 Alh+27ugF1-V	
1845	POOL GP19 CFA+10ugV	
1846	POOL GP20 CFA+10ugV,urea	
1847	POOL GP21 CFA+27ugF1-V	
1848	POOL GP22 CFA+54ugF1-V	İ

50	MMMM	FI	V
Day63	active F1-V	0	0
Day63	active F1-V	40960	81920
Day63	active F1-V	0	1310720
Day63	active F1-V	0	655360
Day63	active F1-V	163840	1310720
Day63	active F1-V	163840	1310720
Day63	active F1-V	o	1280 7
Day63	active F1-V	0	2560 2
Day63	active F1-V	40960	81920
Day63	active F1-V	0	655360
Day63	active F1-V	0	1310720
Day63	active F1-V	163840	655360
Day63	active F1-V	327680	327680

Dato on page 131 in the first devel wedown that the FI-V from proton can in both the 1=1 and V portion of the 1=1-V fusion protein. 19-1 heated animals. If be the probestion slater starte with and the cog2 challeng shows pro If reduce the complexibility REDACTED

Summary REDACTED

· ·	_				F1 Titer	V Tite	er	_
Serum#	Group	Treatment	Bleed Date	Day Post				Change(wells
		Alhydrogel alone	DEDACT	Day63	. 0;	0	. 0	
			REDACT	Day63	40960	81920	163840	ł
		CFA+10ugV	ED	Day63	Oį	1310720	1310720	i
		CFA+10ugV,urea		Day63	0	655360	1310720	
		CFA+27ugF1-V		Day63	163840	1310720	1310720	
		CFA+54ugF1-V		Day63	163840	1310720	1310720	
	POOL GP16			Day63	0	1280	1280	
		Alhydrogel alone		Day63	0	2560	0	
		Alh+27ugF1-V		Day63	40960	81920	163840	
		CFA+10ugV		Day63	, o!	655360	1310720	
		CFA+10ugV.urea		Day63	0	1310720	1310720	
		CFA+27ugF1-V		Day63	163840	655360	1310720	
		CFA+54ugF1-V		Day63	327680	327680	1310720	
	POOL GP23			Day63	0	10240	1280	•
1850	POOL GP24	CFA+27ugF1-V		Day63	163840	163840	1310720	
	1		•	Controls:	i		į	
			:	F1+ Pool	40960	20480	10240	
				831	0	163840	655360	*
			·	Normal Mouse	ا ٥'	5120	U.	

file - Pitt-aerosol data. REDACTED

Aerosol and sc Plague Challenge Expt. (mice)

FOR ACTIVE FILV IMMUNIZATION CHALLIENCE 3 MAR 95

AEROSOL - STEE PAGES 127 - 130

Target

Calculated

Conc./ml

no. CFU/ml

inhaled no. CFU

No. LD50s

suspension 1.75x10e10/ml Prespray

2.8 x 10e10/ml

C092/C12

AGI

1

2

#3

2.5 x 10e8/ml 2.6 x 10e8/ml

3.4 x 10e8/ml

SUBCUTANEOUS -Target

,	1		and the second section of the second	man tanah salah salah salah salah salah salah salah salah salah salah salah salah salah salah salah salah salah		· · · · · · · · · · · · · · · · · · ·			ama rawining benediction only and	i ida oo waxayaa
De	ate: REDACTED	PI:	LTC And	derson					* .	
		Agent:	Plague	e	:	mour me	who			, .
	•	Strain:	_		:		0			
					- 4	27.0				1
		•				32.2				
Anim	nal Model: Mouse				• "	23.4				
Ailiii.	(8) (1)000:000	C12	LD50=1	.1E+05	-	23.7	 			1
VAH- /	(Ave.) : 27.69			•		28.1				
***. /	(7,00,) . 27.00	130				25.6				
	(Ave.): 27.69	7			+			 	· · · · · · · · · · · · · · · · · · ·	-
	· ·					25.6		ļ		
Sev.	female pur T				1	29.0				
OUA.	Torridio p				-					
	cfu	/1	Inhaled	Dose		30.5	 	1		
ΔG	SI/mI AGI aeros		cfu		Strain	3.1				
	E+08 2.50E+09 4.17E			07 94.70	C12	15-4				
	E+08 2.60E+09 4.33E				C12 -	73 - 7	 -	 		-
	E+08 3.40E+09 5.67E				C12	27.695	maren			
	,_,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,		-	·	. ,					
!		-	Summan	DACTED			· · · · · · · ·	 		<u> </u>
		8	summary.							
•										
				REDACTED						
ш .	File: Serumbook	Blood data				E4 TIXED	V TITER			
# 0	Group Treatment GP24 F1-V, day 14 antibody		Bleed day DAY14	Protocol F1-V ANTIBOD	VONTOGEN		40960		 -	
221	GP24 F1-V, day 14 antibody		DAY14	F1-V ANTIBOD			40960		1	
222	GP24 F1-V, day 14 antibody		DAY14	F1-V ANTIBODY			20480			
223	GP24 F1-V, day 14 antibody		DAY14	F1-V ANTIBODY			10240	+	 	+
224	GP24 F1-V, day 14 antibody		DAY14	F1-V ANTIBODY	Y ONTOGEN	Y 40960	640			
225	GP24 F1-V, day 14 antibody	,	DAY14	F1-V ANTIBODY	Y ONTOGEN'	Y 10240	0			İ
226	GP24 F1-V, day 14 antibody		DAY14	F1-V ANTIBODY			10240	 		1
227	GP24 F1-V, day 14 antibody		DAY14	F1-V ANTIBODY			40960			<u> </u>
228	GP24 F1-V, day 14 antibody		DAY14	F1-V ANTIBODY			10240			į
229	GP24 F1-V, day 14 antibody	<u></u>	DAY14	F1-V ANTIBODY			20480			-
			G	eomean(of positive	e values only	31042	13934			
340	GP24 F1-V, day 14 antibody	REDACTED	DAY28	F1-V ANTIBODY	Y ONTOGEN	Y 163840	327680			İ
341	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY			327680	+	-+-	-
342	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY			1280		<u> </u>	_
343	GP24 F1-V, day 14 antibody	1	DAY28	F1-V ANTIBODY		•				
344	GP24 F1-V, day 14 antibody	i i	DAY28	F1-V ANTIBODY	Y ONTOGEN		1	1	 	-
345	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY	Y ONTOGEN'	Y .81920			<u> </u>	
346	GP24 F1-V, day 14 antibody	,	DAY28	F1-V ANTIBODY	Y ONTOGEN'	Y 327680				
347	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY			40960	+		+
348	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY			81920		<u> </u>	
349	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY			163840			
			G	eomean(of positive	e values only!	81920	94101	+	 .	
768	GP24 CFA+27ugF1-V	DED A OFFI	Day63	active F1-V		327680	1310720	1		
769	GP24 CFA+27ugF1-V	REDACTED	Day63	active F1-V		655360	655360			
770	GP24 CFA+27ugF1-V	1	Day63	active F1-V		327680	655360	1	1-1-	1
771	GP24 CFA+27ugF1-V		Day63	active F1-V		327680	1310720	1	 	
772	GP24 CFA+27ugF1-V		Day63	active F1-V		655360	655360	1 .		.
	GP24 CFA+27ugF1-V		Day63	active F1-V		327680	1310720	+	1 1	1
	GP24 CFA+27ugF1-V		Day63	active F1-V		163840	40960	4-1	<u> </u>	<u> </u>
775	GP24 CFA+27ugF1-V	1	Day63	active F1-V		163840	1310720			١.
776	GP24 CFA+27ugF1-V		Day63	active F1-V		655360	1310720	+-+-	1-1-	+
777	GP24 CFA+27ugF1-V	4 4	Day63	active F1-V		Died			<u> </u>	<u> </u>
776	GP24 CFA+27ugF1-V		Day63 Day63	active F1-V	e values onlyl	655360 Died				

		File: Serumbe	ook	REDACTED				
Serum #	Group	Treatment	Bleed date	File Update: Bleed day	Protocol	CHIP#	MISC	V TITER
1688		CFA alone	REDACTED	Day63	active F1-V	1F614E201	2/F1-VB-051	0
1689	GP16	CFA alone	KEBAGTEB	Day63	active F1-V	1F64297C5	8/F1-VB-052	0
1690	GP16	CFA alone		Day63	active F1-V	1F7546317	5/F1-VB-053	0
1691	GP16	CFA alone		Day63	active F1-V	1F626E157	C/F1-VB-054	0
1692	GP16	CFA alone		Day63	active F1-V	1F650F5419	9/F1-VB-055	O
1693	GP16	CFA alone		Day63	active F1-V	1F64075C1	A/F1-VB-056	이
1694	GP16	CFA alone	İ	Day63	active F1-V	1F756A315	1/F1-VB-057	No sample
1695	GP16	CFA alone		Day63	active F1-V	1F6121124	D/F1-VB-058	640
1696	GP16	CFA alone	·	Day63	active F1-V	1F655E405	E/F1-VB-059	이
1697	GP16	CFA alone	_]	Day63	active F1-V	1F65233C1	D/F1-VB-060	<u> </u>
					Ge	omean(of pos	sitive values onlyl) 640
1758	_	CFA alone	REDACTED	Day63	active F1-V		4/F1-VB-121	1280
1759	GP23	CFA alone		Day63	active F1-V		1/F1-VB-122	0
1760	GP23	CFA alone		Day63	active F1-V	1F5F7D107	'5/F1-VB-123	20480
1761	GP23	CFA alone		Day63	active F1-V	1F6507413	4/F1-VB-124	O]
1762	GP23	CFA alone		Day63	active F1-V	1F7172483	6/F1-VB-125	640
1763	GP23	CFA alone		Day63	active F1-V	1F656C4F4	11/F1-VB-126	이
1764	GP23	CFA alone		Day63	active F1-V	1F6018214	8/F1-VB-127	이
1765	GP23	CFA alone	,	Day63	active F1-V	1F6866573	C/F1-VB-128	이
1766	GP23	CFA alone		Day63	active F1-V	1F6278020	5/F1-VB-129	1280
1767	GP23	CFA alone		Day63	active F1-V	1F6508017	3/F1-VB-130	0
								2153

Controls:

Normal Mouse 0 F1+ Pool 5120 Serum 831 327680

of some set. not public, a take colominat or mer libeled late.

Summary REDACTED

			•				
		File: Serumbook		File Update	REDACTED		
Serum #	Group	Treatment	Bleed date		Protocol	F1 TITER	V TITER
2081	GP1 Pool	Alhydrogel only	REDACTED	Day63	ALH-F1-V fusion	0	0
2082	GP2 Pool	ALH+10µgF1	INCONCILO	Day63	ALH-F1-V fusion	81920	
2083	GP3 Pool	ALH+10μgF2urea		Day63	ALH-F1-V fusion	. 81920	
2084	GP4 Pool	ALH+18.5µg F1-V		Day63	ALH-F1-V fusion	81920	163840
2085	GP5 Pool	Alhydrogel only		Day63	ALH-F1-V fusion	0	0
2086	GP6 Pool	ALH+10μgF1		Day63	ALH-F1-V fusion	40960	o
2087	GP7 Pool	ALH+10µgF2urea		Day63	ALH-F1-V fusion	81920	ol
2088	GP8 Pool	ALH+18.5μg F1-V		Day63	ALH-F1-V fusion	81920	327680
2089	GP9 Pool	ALH+37μg F1-V		Day63	ALH-F1-V fusion	163840	163840

REDACTED M pag 127-128 rosed exposure #: 95-079 H Mars Plague react Operator: 56.7 Nov. 1 d. A. M. C. as Key
re operational Check Performed: 66 mg Dry T: 81 West T: 74 Feet, Hern.: 72% P.L. Anderson proced System: No a Otaly Rel Hum. Start Time Agi & Co. Species Dry Wat 69% 102010 35% 80 Mig 80 63 76% 1056 30 C12 506 DE 35% 80 Mice 80 62 25 304 42% Mise 50 25

AEROSOL EXPOSURE SHEET

Date: REDACTED

Aerosol exposure #: 95-032 H

Aerosol	Operator:	RT				_					Agent: PL	AGUE	
Pre-oper	rational Che	ck Perform	ed:		Dry T: &	12 we	a T: 72	Rel, Hum.;	64%		Protocol #:		
Aerosol	System: N	JOSE -	ML	/		_				-	P.L: LTC	ANDE	RSON
System	Flow Rate:	12 LPM	n	_		_							******
Collison	#: B			_									
		Animal	L	Start T		5 min T				r	τ		
Run#	Animal #	Species	Dу	Wel	Rel. Hum.		Wet		Start Time	AGI#	Comments		
1	80 19	MICE	82	70	56%	82	74	70%	1035	50	CIZ	50 L	<u>.D50</u>
ವಿ	20	, rt	32	68	50%	82	74	70%	1101	3	C12	MAX	Imse
											escaped	I min is	55 EXP.
3					HI	B	li	ASH					
									/140				
4	20	MICE	82	68	50%	8,2	75	72%		37A	(092	50 L	-D 50
_ 5	20	MICE	82	69	52%	82	74	72%	1204	フ	C092	MAX	
												A	
									^		movi	GVD	
		•							1) [aΔ	med		
									K, /	RE	EDACTED		
										_			J

Exhibit GA5

Dear Dr Friedlander

CBDE/USAMRIID COLLABORATIVE RESEARCH INTO PROTECTIVE EFFICACY ()F RECOMBINANT V-ANTIGEN AGAINST PARENTERAL AND AEROSOL CHALLENGE WITH YERSINIA PESTIS

As you are aware, CBDE has data to suggest that the V-antigen of the plague causing organism Yersinia pestis, when used as an immimogen, is highly protective against plague. The V-antigen could therefore be a major component of an improved plague vaccine to be developed in the future by CBDE.

You recently indicated to us that USAMRIID wished to collaborate in testing the protective capacity of the V-antigen against parenteral and aerosol challenge with virulent plague. We agreed that such a collaboration would be desirable because it could generate valuable data which would be of benefit to both CBDE and USAMRIID. We therefore decided that the collaboration should, in the future, be the subject of a Project Arrangement under the Memorandum of Understanding between the Secretary of Defense (US) and the Secretary of State for Defence (UK) concerning Technology Research and Development Projects (which is currently still under negotiation).

However, we also agreed that any delay in the collaboration would reduce the benefit of the resulting data, and therefore it would be desirable to commence work in advance of a more formal Project Arrangement.

Accordingly, this letter sets out below the respective duties, rights and responsibilities of each of us under the collaboration, *pro tem*, pending the negotiation of a more comprehensive arrangement:

1. SCOPE OF WORK

- a. CBDE will supply to USAMRIID, for the purposes described in (b), the following:
 - i. 30 mg of recombinant Yersinia pestis V-antigen.
 - ii. Protocols detailing the immunisation route, doses and schedules used at CBDE.
 - iii. Polyclonal antisera raised against the V antigen of Yersinia pestis.
 - 1V. Details of the CBDE challenge route, challenge strain and protection data afforded by the V-antigen vaccine against parenteral challenge with Yersinic pestix.

b. USAMRIID will:

- i. Immunisc groups of animals parenterally with the following:
 - V-antigen in combination with Alhydrogel.

Suggested protocol for the F1-whole V fusion protein.

File: F1-wholeV fusion last update REDACTED

Protocol: B95-01

F1-WV fusion protein immunization and challenge

Investigators: CPT Heath, Dr. Welkos, LTC Anderson, COL Friedlander

Background. CPT David Heath produced and purified a recombinant F1-V fusion protein. The protein is positive by Western blot to F1 and V. Only part of the V-protein was used in the intial F1-V fusion. This F1-V fusion was immunogenic, but was not very efficacious when compared to the whole V-protein (WV) produced by Mauro to protect against F1 minus strains of *Y. pestis*. This is a repeat of part of the intital F1-V protection study using the whole V-protein in the F1-WV fusion. Subcutaneous injection along back of the mouse for immunization.

Purpose: Immunize and challenge mice to check on immunogenicity and protection against the CO92 and C12 strain of Y. pestis by sc and aerosol challenge, $100-Max \ LD_{50}$.

Alhydrogel, 1.3%, from SuperFos. Batch # 2043, Expiration date None, _____µg of AL/dose

Endotoxin level in the F1-WV preparation is ______ U/ml.

Will use Mauro's V which has been urea treated as per the F1-WV procedure. Details in CPT Heath's laboratory notebook.

Immunization Groups: 10 Swiss Webster female mice per group from Harlan Sprague Dawley

Implantable Micro Identification transponders from: BioMeic Data Systems, Inc 255 W. Spring Valley-Ave. Maywod, NJ 07607, 1-800-526-BMDS

Dage

			Dose	
Subcutaneous d	hallenge	Strain	LD ₅₀	# Mice
Group 1	Alhydrogel alone, days 0, 30, sc	C12	100	10 -
Group 2	Alhydrogel + 13.6 μg F1-WV fusion protein day 0, 30, sc	C12	100	102
Group 3	Alhydrogel + 10 μg Mauro-V urea, days 0, 30,sc	C12	Max	10
Group 4	Alhydrogel + 13.6 μg F-1WV fusion protein days 0,30,sc	C12	Max	100
Group 5	Alhydrogel + 27.2 μg F1-WV fusion protein days 0, 30, sc	C12	Max	10×
Group 6	Alhydrogel alone days 0, 30, sc	C12	Max	10/
Group 7	Alhydrogel + 13.6 μg F1-WV fusion protein day 0, 30, sc	CO92	100	10ベ
Group 8	Alhydrogel alone, days 0, 30, sc	CO92	100	10
A				
Aerosol challen				
Group 09	Alhydrogel alone, days 0, 30, sc	C12	50	10 /
Group 10	Alhydrogel + 13.6 μg F1-WV fusion protein day 0, 30, sc	C12	50	10%
Group 11	Alhydrogel + 10 μg Mauro-V urea, days 0, 30, sc	C12	Max	10<
Group 12	Alhydrogel + 13.6 μg F1-WV fusion protein, days 0, 30, sc	C12	Max	107
Group 13	Alhydrogel + 27.2 μg F1-WV fusion protein days 0,30, sc	C12	Max	1.0
Group 14	Alhydrogel alone, days 0, 30, sc	C12	Max	10 ′
Group 15	Alhydrogel + 13.6 μg F1-WV fusion protein days 0, 30, sc	CO92	100	10人
Group 16	Alhydrogel alone days 0, 30, sc	CO92	100	10
Group 17	Alhydrogol + 13.6 µg F1 WV fast prop, days 0, 30, sc	-C12	-Max_	_10~
Group 18	Alhydrogel + 10 μg F1 + 10 μg Mauro's V, days 0, 30, sc	C12	Max	10
Group 19	Greer plague vaccine, days 0, 30, sc	C12	Max	1005
Group 20	Alhydrogel alone, day 0, 30, sc	C12	Max	05
			-	180

10	Group 21	ALH + 13.6 μg F1-WV fusion protein day 0, 30, scantibody response Measure titer at 7.14, 27.57, 96 4.5~		104
. د ر	Group 23	Measure titer at 7,14, 27,57, ALH + 13.6 μg F1-WV fusion protein day 0, 30, sc lung lavage, day 5 ALH + 27.2 μg F1-WV fusion protein day 0, 30, sc antibody response	7 ×	05
	1/ CR 21	Measure titer at 7, 14, 27, 57, 90 Δ·~ ALH + 27.2 μg F1-WV fusion protein day 0, 30, sclung lavage, day 57	یر ۲	05
10 -	Group 25 Group 26	ALH + Mauro-V urea, 10 ug, day 0, 30, scantibody response ALH + Mauro-V urea, 10 ug, day 0, 30, sclung lavage, day 57	×	10
	Group 27	ALH atone, day 0,30, sc Measure titer at 7, 14, 27,57, 90		10
10-	Group29	ALH alone, day 0,30, sc, lung lavage, day 57 ALH alone, for spleen weights 28 day pi	· X Fotal	05 10 05 220

Schedule Groups	1-20	
•	· 	
13Jun95	Arrival of Swiss Webster mice, female 7-8 wks, Harlan Sprague Dawley AA-3 (Barrier)	in
24Jun95	Chipped with BioMedic Data Systems transponders, West	
27Jun95	1st immunization, day 0	
27Jul95	2nd Immunization, day 30	
24Aug95	Bleed to determine prechallenge titers, day 58	
31Aug95	Challenge by aerosol & sc routes, day 65	
28Sep95	Terminal bleed, day 28 pi, titrate spleens# serum #	

Group 21-2	5
13Jun95	Mice arrive
27Jun95	1st immunization, AA-3
11Jul95	Groups 21, 23, 25; Bleed, day 14, AA-3, SERUM#
26Jul95	Groups 21, 23, 25; Bleed, day 29, AA-3, SERUM#
27Jul95	2nd immunization, day
31Aug95	Bleed, day 65, AA-3, Groups SERUM#
	Groups 22, 24, 26, and 28 for serum titer & bronchial lavage #
28Sep95	Group 29 for Spleen weights #
25Sep95	Groups 21, 23, and 25; day 90, serum #

Chip numbers for all groups extra alhydrogel controls

File: 'F1-wholeV fusion

last update REDACTED

Protocol: B95-01

F1-WV fusion protein immunization and challenge

Investigators: CPT Heath, Dr. Welkos, LTC Anderson, COL Friedlander

Background. CPT David Heath produced and purified a recombinant F1-V fusion protein. The protein is positive by Western blot to F1 and V. Only part of the V-protein was used in the intial F1-V fusion. This F1-V fusion was immunogenic, but was not very efficacious when compared to the whole V-protein (WV) produced by Mauro to protect against F1 minus strains of Y. pestis. This is a repeat of part of the intital F1-V protection study using the whole V-protein in the F1-WV fusion. Subcutaneous injection along back of the mouse for immunization.

Purpose: Immunize and challenge mice to check on immunogenicity and protection against the CO92 and C12 strain of Y. pestis by sc and aerosol challenge, 100-Max LD₅₀.

Alhydrogel, 1.3%, from SuperFos. Batch # 2043, Expiration date None, 0.1857 4g of AL/dose

Endotoxin level in the F1-WV preparation is _____ U/ml.

Will use Mauro's V which has been urea treated as per the F1-WV procedure. Details in CPT Heath's laboratory notebook.

Immunization Groups: 10 Swiss Webster female mice per group from Harlan Sprague Dawley

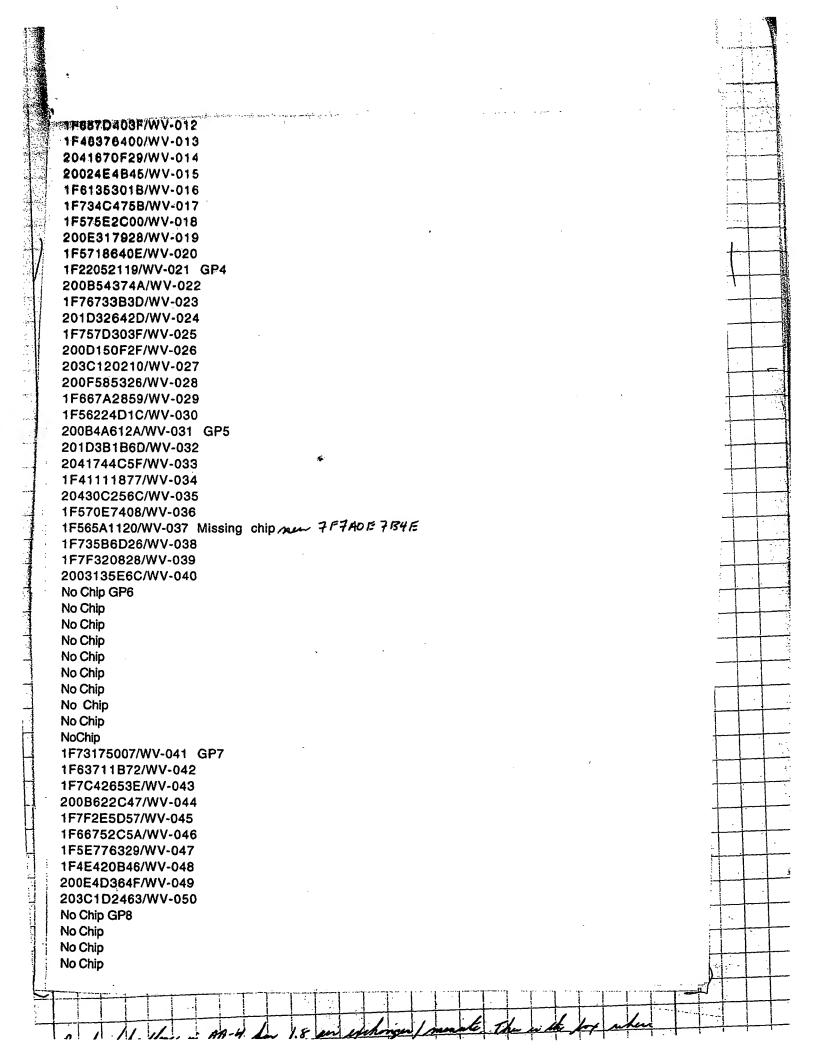
Implantable Micro Identification transponders from: BioMeic Data Systems, Inc 255 W. Spring Valley Ave. Maywod, NJ 07607, 1-800-526-BMDS

			Dose	
Subcutaneous	challenge	Strain	LD ₅₀	# Mice
Group 1	Alhydrogel alone, days 0, 30, sc	C12	100	10 NC
Group 2	Alhydrogel + 13.6 μg F1-WV fusion protein day 0, 30, sc	C12	100	10
Group 3	Alhydrogel + 10 μg Mauro-V urea, days 0, 30,sc	C12	Max	10
Group 4	Alhydrogel + 13.6 μg F-1WV fusion protein days 0,30,sc	C12	Max	10
Group 5	Alhydrogel + 27.2 μg F1-WV fusion protein days 0, 30, sc	C12	Max	10
Group 6	Alhydrogel alone days 0, 30, sc	C12	Max	10 NC
Group 7	Alhydrogel + 13.6 μg F1-WV fusion protein day 0, 30, sc	CO92	100	10
Group 8	Alhydrogel alone, days 0, 30, sc	CO92	100	10 NC
Aerosol challe	enge			
Group 09	Alhydrogel alone, days 0, 30, sc	C12	50	10)
Group 10	Alhydrogel + 13.6 μg F1-WV fusion protein day 0, 30, sc	C12	50	10)
Group 11	Alhydrogel + 10 μg Mauro-V urea, days 0, 30, sc	C12	Max	10
Group 12	Alhydrogel + 13.6 μg F1-WV fusion protein, days 0, 30, sc	C12	Max	10
Group 13	Alhydrogel + 27.2 µg F1-WV fusion protein days 0,30, sc	C12	Max	10
Group 14	Alhydrogel alone, days 0, 30, sc	C12	Max	10
Group 15	Alhydrogel + 13.6 µg F1-WV fusion protein days 0, 30, sc	CO92	100	10
Group 16	Alhydrogel alone days 0, 30, sc	CO92	100	10
Group 17	Alhydrogel + 10 μg F1 + 10 μg Mauro's V, days 0, 30, sc	C12	Max	10
Group 18	Greer plague vaccine, days 0, 30, sc	C12	Max	09 NC
Group 19	Alhydrogel alone, day 0, 30, sc 40T/0/3w2	C12	Max	05
Group 20	ALH + 13.6 μg F1-WV fusion protein day 0, 30, scantibody Measure titer at 7,14, 27,57, lung lavage day 57	response		10

Puntoy bordline in AA-4 har 1.8 air exchanger menate. The in the box

	Managera titar at 77 14 27 57; lung layang day 57	
Group 22	Measure titer at 文, 14, 27, 57; lung lavage day 57 ALH + Mauro-V urea, 10 ug, day 0, 30, scantibody response	10
Group 23	ALH alone, day 0,30, sc	10
: Group 20	Measure titer at ✓ 14, 27,57; lung lavage day 57	10 :
Group 24	Greer plague vaccine days 0, 30, sc	
	Measure titer at X, 14, 27, 57; lung lavage day 57 Chipped	05
Group 25	Alhydrogel + 10 µg F1 + 10 µg Mauro's V, days 0, 30, sc	<u>07</u>
<i>7</i>	Measure titer at X 14, 27, 57; lung lavage day 57, Mice rec'd 14Jun95	<u></u>
	Total	232
Schedule	4	
Groups	1-19	7:
13Jun95	Arrival of Swiss Webster mice, female 7-8 wks, Harlan Sprague Dawley in	·
	AA-3 (Barrier)	
24Jun95	Chipped with BioMedic Data Systems transponders, West	İ
27Jun95	1st immunization, day 0, Zimmerman, West, Giunanzio, Archer, Anderson	<u> </u>
25Jul95	2nd Immunization, day 28, Mil, wat, Thurage Unk, Order	
24Aug95	Bleed to determine prechallenge titers, day 58 5664 - 5847	
0 8 Sep95	Challenge by aerosol & sc routes, day 55, AA-3-5849-5859	+
28Se p95 ∞ ∞ σ	Terminal bleed, day 28 pi, titrate spleens# 10030 - 11094 serum # 60	82 -6187
	21 MARICANA SHAMBLIN	
Group 2 13Jun95	0-23	+
27Jun95	Mice arrive 1st immunization, AA-3	-
11Jul95	Groups 20, 21, 22; Bleed, day 14, AA-3, SERUM# 4542 - 4593	
26Jul95	Groups 20, 21, 22; Bleed, day 29, AA-3, SERUM# 4807-488 GIATTA	
25Jul95	2nd immunization, day28, new barrier, AR-5	, ,
0 4 Sep95	Bleed, day 65, AA-3, Groups 24,25 SERUM# 5848 - 5899 GIWAL 214 F	AALL
-4	Groups 20, 21, 22, and 23 for serum titer & bronchial lavage #5 900 - 395	-1
0 \$Sep95	Group 23 for Spleen weights # /00/0 - /00/9	
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Use scanner to check chip number of dead mice Mark number of animals alive in each cage re = mouse has been rechipped

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3 scanner to check chip number of dead mice rk number of animals alive in each cage

= mouse has been rechipped

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Discard dead animals

Use scanner to check chip number of dead mice Mark number of animals alive in each cage re = mouse has been rechipped

11.

File: F1-V,longterm

Last update: REDACTED

Prótocol: B95-01

Investigators: Anderson/Heath/Welkos/Friedlander

Background: F1-WV and F1 and V in combination have been shown to protect against challenges of CO92 and C12 with a two dose schedule (0 and 30). The long-term decay of the antibody response to the initial immunization and length of protection from a single immunization is currently not known.

Purpose: To examine the decay of the antibody response to an initial immunization, protection afford over time to an initial immunization to indicate the optimum time for the 2nd immunization for an aerosol challenge. Titers to F1 and V will be determined. Mice will be challenged with 50-100 LD₅₀ CO92, aerosol challenge.

In the below challenge groups, when protection falls to zero, the remaining groups will be booster and challenged 2 weeks post-boost.

Immunogens: EcF1s, Mauro's V, and F1-WV all essentially endotoxin free from Dr. Heath.

Mice: Swiss Webster (Hsd:ND4) female mice per group from Harlan Sprague Dawley, Inc. Indianapolis, Indiana (317) 894-7521.

Alhydrogel: 1.3%, from SuperFos. Batch # 2043, Expiration date None, 0.1857 mag of AL/dose

IMI-1000, Implantable micro identification transponders, BioMedic Data Systems, Inc., 255 W. Spring Valley Ave. Maywood, NJ 07607, Tel 201 587-8300.

Plague USP, Lot #______, Expiration Date REDACTED, Greer Laboratories, Inc. P.O Box 800 Lenoir, NC 28645-0800

Group 1 A 1 B	Treatment 10μF1+20μgMauro-V day0	Challenge Day 14	#of mice 10 05 10 05 10 05 10 05 10 line A- /0/46-10/60 10 20 10 10 10 10 10 10 10 10 10 10 10 10 10
2 A 2B	30μgF1-WV	14	10 Serum # 1990 - 7039 - por or page 1
3	Plague USP	14	10 \ line 1 /0/46 - 10/60
4	Alhydrogel alone	14	10) see 4 7820 - 7837 olay 25 P1 T
5 A 5B	10μF1+20μgMauro-V day0	42	10 05 10 05 10 10 10 10 10 10 10 10 10 10
6 A 6B	30µgF1-WV	42	10 \ 2676-7725 per 10091 day 81 per 1 habeley
7	Plague USP	42	10 \ lessen 10/60
8	Alhydrogel alone	42	10) man 4 5004 - 5035 M301
9 A 9B	10μF1+20μgMauro-V day0	.98 119	10 05
10A 10B	30μgF1-WV	98 119	10 10 10 10 10 10 10 10 10 10
11	Plague USP	93 119	10 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1
12	Alhydrogel alone	98119	10) him " 10257-1021

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10μF1+20μgMauro-V day0
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13A
13B
                                                    05
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       30µgF1-WV
14A
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14B
15
       Plague USP
                                                    10
16
       Alhydrogel alone
                                                    10/
17A
       10μF1+20μgMauro-V day0
                                                    10 serial bleeds on challenge days,
17B
                                                    05
18A
       30µgF1-WV
                                                    10
                                                           14, 42, 93, and
                                                        day 14 serum # 7040 - 3077
18B
                                                    05
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19
       Plague USP
                                                    10
20
                                                    <u>10</u>
       Alhydrogel alone
                                                           93
                                            Total
                                                  250
Schedule
17Oct95
              Mice arrive B412
              Mice chipped B412, Plumtree, Zimmerman
25Oct95
              Mice immunized day 0, Plumtree, Zimmerman
31Oct95
              Bleed 1st challenge group, day 7 serum #
07Nov95
               1st challenge group, day 14, 100 LD<sub>50</sub> aerosol challenge, CO92
14Nov95
14Nov95
              Bleed serial bleed group, day 14
05Dec95
              Bleed 2nd challenge group, day 35
12Dec95
              2nd challenge group, day42, 100 LD<sub>50</sub> aerosol challenge, CO92
              Bleed serial bleed group, day 42
12Dec95
                                                  PLIMTRIES SAMMBUN, AMORIASO ~
              Bleed 3rd challenge group, 86 PAY
25Jan96
01Feb96
              3rd challenge group, day 93, 100 LD<sub>50</sub> aerosol challenge, CO92
              Bleed serial bleed group, day 93
01Feb96
              Bleed 4th challenge group, day
 95
              4th challenge group, day, 100 LD<sub>50</sub> aerosol challenge, CO92
 ____96
  96
              Bleed serial bleed group, day
Chips Numbers
22254D6722/LT-001 GP1A
22254B4164/LT-002
221D487E7E/LT-003
221D705617/LT-004
22213D0B42/LT-005
222122186F/LT-006
22223E1239/LT-007
2221493775/LT-008
221D670720/LT-009
22213D5220/LT-010
222233166C/LT-011 GP2A
2227765159/LT-012
221D6C477D/LT-013
2222485A77/LT-014
221A2B093D/LT-015
221D6B7917/LT-016
221D686E17/LT-017
2222411328/LT-018
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22225C6018/LT-019 221D630D5E/LT-020 221D4E2460/LT-021 GP3

22280F622F/LT-022 221D570056/LT-023 221D552A40/LT-024 22223C1767/LT-025 2221534945/LT-026 22277D234D/LT-027 221D660734/LT-028 221B495727/LT-029 2225577D72/LT-030 221B45423B/LT-031 GP4 221B4C6576/LT-032 221D5B6077/LT-033 2222405171/LT-034 22277E167A/LT-035 221D682963/LT-036 222251294B/LT-037 221B464652/LT-038 221D605813/LT-039 221D762A24/LT-040 22224F0E15/LT-041 GP5A 22252A534C/LT-042 221D627877/LT-043 22214B1228/LT-044 2219346F3E/LT-045 221D4F3856/LT-046 2227545533/LT-047 22214E1004/LT-048 2222455747/LT-049 221B366372/LT-050 2227624147/LT-051 GP6A 22217E1A0D/LT-052 2228195A13/LT-053 2222585550/LT-054 2222390E64/LT-055 22214D1F3C/LT-056 2225405710/LT-057 221D713C0B/LT-058 22225B030A/LT-059 221D645754/LT-060 221B502C5D/LT-061 GP7 22276D2115/LT-062 222142305C/LT-063 22215D0B34/LT-064 2222375E5F/LT-065 2228166739/LT-066 221D6D0B4C/LT-067 2222576F77/LT-068 222155635F/LT-069 2222480418/LT-070 2221464260/LT-071 GP8 2222444E48/LT-072 221D72505E/LT-073 221D6D0524/LT-074 222243493A/LT-075

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221D524871/LT-183

221D742C6D/LT-184 221B471829/LT-185 221D59352E/LT-186 2222476C0C/LT-187 2222511C0F/LT-188 22277F7C66/LT-189 221D4B422C/LT-190 221D6B4536/LT-191 GP20 2221710D56/LT-192 22281C2918/LT-193 221D5C6E60/LT-194 2221662F72/LT-195 221D5E3068/LT-196 22223A6E6D/LT-197 22277C0B1C/LT-198 221D6C3C28/LT-199 2228045804/LT-200 2221463F1B/LT-201 GP1B 22213F0074/LT-202 22215A2E71/LT-203 22223B1136/LT-204 22215A7309/LT-205 221D6B4C53/LT-206 GP2B 221D49253C/LT-207 221D710C76/LT-208 22273F017C/LT-209 22217E5E6E/LT-210 221D494F34/LT-211 GP5B 22217B022D/LT-212 2221386837/LT-213 2228057813/LT-214 222142434F/LT-215 2222323813/LT-216 GP6B 222121405D/LT-217 2221755864/LT-218 2225527970/LT-219 221D56403B/LT-220 221D681542/LT-221 GP9B 22280E312B/LT-222 221D584D78/LT-223 221D4C4067/LT-224 22212A2966/LT-225 222124374C/LT-226 GP10B 2222586303/LT-227 2222371808/LT-228 2222443D5B/LT-229 22213B707C/LT-230 221D737744/LT-231 GP13B 2221313035/LT-232 2222392702/LT-233 221D73716A/LT-234 221D527D35/LT-235 221B2E7B71/LT-236 GP14B 221D484645/LT-237

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File: Alhydrogel concentration Last updated: REDACTED

Anderson/Heath/Welkos/Friedlander

Background. The allowable AI content in a human vaccine is 0.85 mg/dose as determined by assay (21 CFR 610.15(1)). However, the lowest possible dose of AI should be used which maintains an adequate adjuvant response with the EcF1c and V immunogens. A dose response for Alhydrogel has not been done with a combination of F1 and V. Therefore this experiment will examine a range of concentrations of AL which will be used with a constant amount of EcF1c and V. EcF1c (60 EU/ml) and V-His tag (preparation are essentially endotoxin free. The level of endotoxin in the thrombin treated V preparation without the His tag is 49 EU/ml.

Compare the antibody response to V with F1=WV protein with and without alhydrogel. When F1 + V is used to immunize without alhydrogel, there was not antibody response to V. This will be done with F1-WV in order to determine whether F1 is contributing anything to protection with the F1-WV protein. F1-WV purified by Ni++ and ran through a Sartorius Q15 filter using 10mMtris, ph 7.6, 0.5mM EDTA + 0.5 MNaCl for elution. See Heath's note of 5-6 Dec. Endotoxin _____ EU/ml.

Mice: Swiss Webster (Hsd:ND4) female mice from Harlan Sprague Dawley, Inc. Indianapolis, Indiana (317) 894-7521.

1.3% Alhydrogel (Aluminium Hydroxide Gel Adjuvant): Al_2O_3 (1.3%) equivalent to $Al(OH)_3$ (2.0%), from SuperFos Biosector a/s, Frydeniundsvej 30, DK-2950 Vedbaek, Denmark. Batch # 2043, Expiration date None: U.S. supplier - Accurate Chemical & Scientific Corp, 300 Shames Drive, Westbury, NY 11590, Tel (516) 333-2221, Fax (516) 997-4948.

AI = 13 O = 8 H = 1, $AI(OH)_3 = 40$ molecular weight

Current procedure for adsorption of F1 and V to Alhydrogel: 1.0ml Alhydrogel brought to 7.0 ml 2% Al(OH)₃ = 20 mg/ml

20mg/ml)(1.0ml) = (x)(7.0ml final volume) x = 2.857 mg Al(OH)₃/ml (2.857mg/ml)(0.2ml dose) = 0.57142 mg Al(OH)₃

AL is 0.325% of AI(OH)3

0.235% of 2.857mg = 0.1857 mg of AL/0.2ml dose which the mouse receives

Current dose of AI which has been used through out the mouse experiments is 0.1857mg. Try two other doses each 75% of the former GP1 = 0.1857mg (standard amount of AL), GP2 = 0.1393mg, GP3 = 0.1045mg

IMI-1000, Implantable micro identification transponders, BioMedic Data Systems, Inc., 255 W. Spring Valley Ave. Maywood, NJ 07607, Tel 201 587-8300.

Plague USP, Lot # 1128X1, Expiration Date REDACTED, Greer Laboratories, Inc. P.O Box 800 Lenoir, NC 28645-0800,

Groups	Treatment	V-HIS Tag	Strain	#Mice
000A	30μgF1-WV, μο AL	Yes	CO92	10
000B	30μgF1-WV , Ma AL	Yes	CO92	05
00A	30μgF1-WV+0.19Al	Yes	CO92	10
00B	30μgF1-WV+0.19Al	Yes	CO92	05
0 A	10μgF1+20μgV+0.19Al	No	CO92	10
9B	10μgF1+20μgV+0.19AI	No	CO92	1-0

The state of the s	1 A	10μgF1+20μgV+0.19Al	Yes	CO92	10		
	1B	10μgF1+20μgV+0.19Al	Yes	CO92	10	\$	
- E	2À	10μgF1+20μgV+0.14Al	Yes	CO92	10		
	2B	10μgF1+20μgV+0.14Al	Yes	CO92	10		•
	3 A	10μgF1+20μgV+0.10Al	Yes	CO92	10		
#	3B	10μgF1+20μgV+0.10Al	Yes	CO92	10		
-#	4	0.19Alhydrogel		CO92	10		
1	5	0.19Alhydrogel		CO92	10		
	6	0.14Alhydrogel		CO92	10		
	7	0.10Alhydrogel		CO92	10		
	, 8	10μgF1+20μgV, No Alh	Yes	CO92	10		
	9	10μgF1+20μgV, No Alh	Yes	CO92	10		
	10	Plague USP (Greer)sc		CO92	10		
	11	Plague USP (Greer)sc		CO92	10		
	12	No treatment		CO92	10		
	·13	Plague USP (Greer) im		CO92	<u>10</u>		
		, ,		•	Total 210		
•					_		
	Date	Procedure					
	09Nov95	Arrival of mice in B412 at 7-8	weeks	of age			
	16Nov95	Chipped mice in B412, SGT Z	immerm	an/Plumtre	е	•	
•	06Dec95	Day 0, immunization Anderson	n/Shamb	lin	•		
	12Jan96	Day37, bleed prior to challeng	e, serun	n # 8045 -	-8254 Paum mil	is, Andiazon SHA	MPLIN
	19Jan96	Day44, challenge Aminason, 611	INGS FIN	IN ANIMA	prom + BINT	151 3 MINS	
	15Feb96	Day 28 pi, terminal bleed, sple	en rem	oval			
		serum #		spleen #			
	Chip numbers					•	
	221D682232/A	LH-001 GP000A					
	2225353065/	ALH-002				•	
	222543363A/	ALH-003					
	2227471E04/A	NLH-004					
	2221462517/A	NLH-005					
	221D63634C/	ALH-006					
	221D645F12/A	NLH-007		•			
	221B2F0113/A	LH-008					
•	222537362C/A	ALH-009					
	221B4D572B/	ALH-010				•	
	221D3A1A63/A	LH-011 GP000B	•	,			
	2227593C63/A	NLH-012					
	22213E5370/A	LH-013					
	2228111E27/A	LH-014					
	221B4D060F/A	LH-015					
	2221291C51/A	LH-021 GP00A					
	22217A1916/A	NLH-022					
	22215A4825/A	NLH-023					
	22213D0C4F/A	•					
	2221661921/A	LH-025					
	2221501E3C/A						
	221D62561C/A						
	2221796313/A						
	22216A4521/A		•				
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	221D5B165D/A 2225535315/A						

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REDACTED 645 1A +1B long FX + 20 mg hV in Fac final volume hV= 13 EW/ml 496 al FI (705 mg/ms) + 129 ml hv (514 mg/ms) + 375 ml PBS + IML ALHADESTE, KOCK BU @ 40C - Spin 2000 rpm, 5 min. remove 2, 10 ul aliquoto for BCA Assay of absorption added P85 to 7 at final volume 6 KBS 1A + 2B same to 1A + 1B above but used 750 al ALHYDCOG additional 250 ul PBS 6KIS 34 + 3B same as 1A + 1B store but used 562, In ACHIDEONIC + 437.5 L PBS to equal (me then same is IA+1B Held fac sufferen to lac 185 + Local DV. 6RP 4 +5 had volume to 7 mc + 250 ml 155 + /mc PBS & HLPS 6 alled 750 ml satyons rocked DN @ 4ºC. added 185 to Face hood volume. added 562. Sal Actyp 2062 + 437.5 1855 then Inc 185 GRP 7 a rold Du @ 452 -abled PSS to Fare find orland 496 al FI + 129 al hV + 1.375 mc PBS Y weeked 8V. RP 819 Philed The 111.1 1855

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Discard dead animals

Check chip number of dead mice with scanner Mark number of animals alive in each cage LOST after Chip # means the chip has fallen out

FEAR- MESSE CHAIL OUT OF HELDIN PURIL CHALLENGE

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For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

Discard dead animals

Check chip number of dead mice with scanner / Y / - No / SUME MUUSIE MELLISUIM FOUNTEYPISURE

Mark number of animals alive in each cage

LOST after Chip # means the chip has fallen out

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Control and Control March Control and Control

ELISA Summary: Protocol: F1-V Long Term (Day 7)

Protocol: F1-V Long Term (Day 35)

		v	ELISA DATE:	30 JAN 96	FI	ELISA DATE:	25 JAN 96
Plate	Serum	Group	Treatment	V TITER	V Geomean	F1 TITER	F1 Geomean
14	6990	GP1A10ugF1	+20ugMaurV	320	485	2,560	2,681
18	6991	GP1A10ugF1	+20ugMaurV	640		1,280	
2A _	6992	GP1A10ugF1	+20ugMaurV	2,580		10,240	
28	6993	GP1A10ugF1	+20ugMaurV	640	ľ	2,560	l l
34	6994	GP1A10ugF1	+20ugMaurV	640		2,560	l.
38	6995	GP1A10ugF1	+20ugMaurV	320		2,560]
48	6996	GP1A10ugF1	+20ugMaurV	320		2,560	
4B	6997	GP1A10ugF1	+20ugMaurV	320		2,560	
5A	6998	GP1A10ugF1	+20ugMaurV	640		5,120	
58	6999	GP1A10ugF1	+20ugMaurV	1,280	i	5,120	•
21A	7030	GP1B10ugF1	+2QugMaurV	320		1,280	
21B	7031	GP1B10ugF1	+20ugMaurV	320	1	320	
22A	7032	GP1B10ugF1	+20ugMaurV	320		5,120	
22B	7033	GP1B1CugF1	+20ugMaurV	320		2,560	
23A	7034	GP1B10ugF1	+20ugMaurV	320		5,120	
BA	7000	GP2A		320	557	2,560	3,225
68	7001	GP2A	30ugF1-WV	640		1,280	٠,٠
7A	7002		30ugF1-WV	640		5,120	l
7B	7003		30ugF1-WV	2,560		2,560	
84	7004		30ugF1-WV	640		5,120	
88	7005		30ugF1-WV	640		1.280	- 1
9A	7006		30ugF1-WV	320		2,560	
98	7007		30ugF1-WV	320		5,120	1
10A	7008	· GP2A	30ugF1-WV	1,280		5,120	
108	7009		30ugF1-WV	640		5,120	
238	7035		30ugF1-WV	640		10,240	
24A	7036		30ugF1-WV	320		5,120	
24B	7037		30ugF1-WV	320		10,240	I
25A	7038		30ugF1-WV	320		2,560	I
258	7039	GP2B	30ugF1-WV	640		320	
11A	7010		PlagueUSP	320	343	2,560	2,389
118	7011		PlagueUSP	320	0.0	5,120	
12A	7012		PlaqueUSP	320		2,560	. !
12B	7013		PlaqueUSP	320		640	i
13A	7014		PlagueUSP	320		5,120	- 1
138	7015		PlagueUSP	640		640	
144	7016		PlagueUSP	320		2,560	I
14B	7017		PlaqueUSP	320		2,560	
15A	7018		PlaqueUSP	320		2,560	I
158	7019		PlagueUSP	320		5,120	I
18A	7020		alhydro alone	320	343	320	320
168	7021		alhydro alone	320		320	320
17A	7022		athydro alone	320		320	
178	7023		athydro alone	320		320	
18A	7024		alhydro alone	320		320	1
18B	7025		alhydro alone	320		320	
19A	7026		alhydro alone	320		320	1
19B	7027		elhydro alone	640		320	I
20A	7028	GP4	alhydro alone	320		320	I
20B	7029		alhydro alone	320		320	
338	FIN	POOL (+ CONT		327,680		81,920	
				3E1,000		01,520	

			ELISA DATE:			ELISA DATE:	6 FEB 96
Plate	Serum	Group	Treatment	V TITER		F1 TITER	F1 Geomean
1A	7676	GP5A10ugF1		1,310,720	1,040,319	40,960	25,803
1B	7677	GP5A10ugF1		1,310,720		40.960	
2A	7678	GP5A10ugF1		1,310,720		40,960	
28	7679	GP5A10ugF1		1,310,720		40,960	
3A	7680	GP5A10ugF1		1,310,720		20,480	
38	7681	GP5A10ugF1		327,680		40,960	
4A	7682	GP5A10ugF1		1,310,720		20,480	
4B	7683	GP5A10ugF1		1,310,720		20,480	
5A	7684	GP5A10ugF1		1,310,720		40,960	
58	7685	GP5A10ugF1		1,310,720		81,920	
6A	7686	GP5B10ugF1		1,310,720	1	10,240	
6B	7687	GP5B10ugF1		1,310,720		10,240	
7A	7688	GP5B10ugF1		1,310,720	1	20,480	
78	7689	GP5B10ugF1		1,310,720		10,240	
8A	7690	GP5B10ugF1		163,840		20,480	
8B	7691	GP6A ALH+		655,380	948,482	81,920	9,336
9A	7692	GP6A ALH+		1,310,720	340,406	5,120	0,000
98	7693	GP6A ALH+		655,360		20,480	
10A	7694	GP6A ALH+		1,310,720		10,240	
108	7695	GP6A ALH+		1,310,720		10,240	
11A	7696	GP8A ALH+		163,840	ļ	2,560	i
11B	7697	GP6A ALH+		655,360	_	2,560	
12A	7698	GP6A ALH+		1,310,720		20,480	
128	7699	GP6A ALH+		655,360		20,480	
13A	7700	GP6A ALH+		1,310,720		5,120	
138	7701	GP6B ALH+		1,310,720		20,480	
14A	7702	GP68 ALH+		1,310,720		20,480	
14B	7703	GP6B ALH+		1,310,720		2,560	
15A	7704	GP6B ALH+		1,310,720		10,240	
15B	7705	GP6B ALH+		1,310,720		2,560	
16A	7706		Plaque USP	320	735	320	4,777
168	7707			1,280	/ "	20,480	-,,,,,
17A	7708		Plague USP	640	1 /	10,240	
17B			Plague USP Plague USP	540	17	20,480	
18A	7709 7710		Plaque USP	1,280		1,280	
188	7711			640	\sim	1,280	•
19A	7712		Plague USP	64C	11)	5,120	
198	7713		Plague USP	320	11 /	2,560	
20A	7714		Ptague USP	1,280	I <i>I I</i>	5,120	
20B	7715		Plaque USP	. 1,280	<i>//</i> /	40,960	
			Plague USP		100		320
21A	7716		Alhydrogel	640	485	320 320	320
21B 22A	7717		Athydrogel	640	171	320	
	7718		Alhydrogel	320	11 1	320	
228	7719		Alhydrogel	640	IU I	320	
23A	7720		Alhydrogei	640	174	320	
23B	7721		Alhydrogel	320	[7]	320	
24A	7722		Alhydrogel	320	11 }	320	
24B	7723		Alhydrogel	1,280			
25A 25B	7724		Athydrogel	320	V I	320 320	
	7725		Athydrogel	320	<u> </u>		
338	F1/V	POOL (+ CONT	HOL	655,360		81,920	

<u>.</u>			
Mouse weights	for aerosol run o	f 23Feb	96
24.9			
30.9	average wt (gm)	29.98	
32.5	Cage 9A		
32.4			
29.5			
28.8			
28.2	•		
30.1			/
34.3			
28.2			
	,		

Ig Subclass ELISA response to vaccination with Yersinia pestis V and F1 antigens (continued)

Plate setup:

Same as 21 FEB 96, except test samples: (9) for V satigen only.

Procedure:

Same as 21 FEB 96, except:

*Second antibodies (goat anti-mouse IgG1, IgG2a, and IgG2b) dilute
1:3000 (decreased from 1:2000).

*Conjugate dilution decreased to 1:3000 (from 1:2000)....

Results: see separate pages.

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BAIT-V	day 58 ports	managadan	•					
<u>Sample</u>	Treatment	IaG(H+L)	laG1	lqG2a	lgG2b			
2041	Alhydrogel	320	320	320	640			
2042	Alhydrogel	320	320	320	320			
2043	Alhydrogei	320	320	320	320			
2044	Alhydrogel	320	320	320	320			
2045	,, _ , _, _	320	320	320	320			
2046	Alhydrogel	320	320	320	320			
2047	Alhydroget	320	320	320	320			
2048	Alhydrogel	320	320	320	320			
2049	Alhydrogel	320	320	320	320			
2061	/ Alh+V	640.000	655,360	20,480	31,920			
2062	Alh+V	320.000	1.310,720	81.920	10,960			
2063	(Alh+V	540,000	1.310,720	40.960	31,920			
2064	Alh+V	320,000	355,360	40.960	40,900			
2065	Alh+V	540,000	335,360	31,920	10.960			
2066	√ Alh+V	340,000	327,680	≟0.960	20,480			
2067	1	:.280,000	1.310,720	31.920	31 320-0			
2058	/ Alh+V	1.230,000	1.310.720	40.960	160,8402			
2069	. Alh+V	540,000	1.310.720	20.480	11 420 A			

	exposure #			-	Location	<u></u> 4/	0-4 H	_			Date: 23 F.(9)
	Operator:			- /							Agent: Diagra (05)
	rational Che			<u> </u>	Dry T: יך	(v.	1 T: 63	Rel. Hum.:	63%	_	Protocol #:
	re System T			-							P.I.: COL Byrn
System	Flow Rate:	12.0 4) M	_			Timer Ch	ieck: 🗸	Pre 🗸	Post	LTC Andrison
Collison	#: A	~					Start time	e 930	14	100	-
Panel #:	3		_					0935		07	-
Electron	ic Flow Met	or#: Æ	6479					- 0777		<u>U/</u>	- '
		Animal									-
Run #	Animal #	Species	Dry	Start T Wet	Rel. Hum.	Dry	5 min T Wet	Rel. Hum.	Start Time		
1	35		71	63	u3%					$\overline{}$	Comments
L		Mir	\vdash	62	7	71	67	80%	1001	52	
<u> </u>	15	Mire	73	63	58 %	73	68	78.5%	1030	1163	# Councit Lower Humbity , becau
3_	15	mira	73	63	58%	73	68	78.5%	1054	361	all air bubbles to second
4	25	mile	73	63	58%	73	68	78.5%		$\overline{}$	am lovered off (But)
5_	23	Mica	74	63	55%	74	65	68%	1152	31	
ι	35	Mica	75	64	57%	15	68	70%		360	
7	15	Mice	75	63	51%	74	66	68%	1371	74	* Ont mouse dad on arrive
				"	71.0	<u> </u>	106	6010			· ////
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Antibiotic Treatment / Vaccine Challenge

Date:

23-Feb

PI:

COL Byrne / LTC Anderson

Agent:

Plague

Strain:

CO92

Animal Model: Mouse Strain: Swiss Webster

Wt: (Ave.) : 20g / 29.98g

CO92

LD50=2.1E+04

for um 647, andress

Sex: female

			cfu/l		Inhaled Dose		
Run #	AGI/ml	AGI	aerosol	MV	cfu	LD50s	
1	3.50E+07	3.50E+08	5.83E+06	0.02	1.17E+06	55.56	CO92
2	3.80E+07	3.80E+08	6.33E+06	0.02	1.27E+06	60.32	CO92
3	5.10E+07	5.10E+08	8.50E+06	0.02	1.70E+06	80.95	CO92
4	4.00E+07	4.00E+08	6.67E+06	0.02	1.33E+06	63.49	CO92
5	8.10E+06	8.10E+07	1.35E+06	0.02	2.70E+05	12.86	CO92
6	1.70E+07	1.70E+08	2.83E+06	0.027	7.65E+05	36.43	CO92
7	1.60E+07	1.60E+08	2.67E+06	0.027	7.20E+05	34.29	CO92

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Date: 23 Noteboo		739	Pe	OLECI	: Act	ve	<u>-1-</u>	wv	<u>, F1</u>	+4	ron	i ex	stu	dy I)a⊭	419	Pos	<u>stimn</u>	nuni.	zaso		_		<u> </u>		-		^	_	5		-	_	
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Route: a Animal si		wiss			s sho	own) be		rival	10	5°0; 1/17/	95	<u>7 - 3</u>	6 8w	ke.		Tvz	odor	· Ha	don	`	gue (i			la.						7		
Month - F	eb D	ay -			25	26	27	28	3 2 9	1		Ĭ		E		7		9	10	[11]	12	13 1	4 1	5 1	61.17	118	x: fe	20	2	122	2 2 :	3	·	
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9A	CO92		1	1	1	Т	1	$\dagger \tau$	$\dagger \tau$	ti	+	1	+	h	$\dagger \tau$	1	╁	1	1	1	┪	┰┼	, 1	+,	+,	١,	1:	-	+	+	╀	221D54560	C# T 001	
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	1000		V Summary	The day is	
	TOCOL: V				V antigen TITER
Plate			Treatment	Bleed	lgG(H+L)
1A	8288	-	F1+V	+DAY86	81,920
<u>1B</u>	8289		F1+V	+DAY86	327,680
2A	8290		F1+V	+DAY86	40,960
28	8291	GP9A	F1+V	+DAY86	NO SERUI
3A	8292		F1+V	+DAY86	163,840
38	8293	GP9A	F1+V	+DAY86	327,680
4A	8294		F1+V	+DAY86	327,680
48_	8295	GP9A	F1+V	+DAY86	655,360
5A	8296	GP9A	F1+V	+DAY86	40,960
58	8297		F1+V	+DAY86	655,360
3A	8298	GP98	F1+V	+DAY86	327,680
38	8299	GP9B	F1+V	+DAY86	327,680
/A	8300	GP9B	F1+V	+DAY86	81,920
8	8301	GP98	F1+V	+DAY86	655,360
M_	8302	GP9B	F1+V	+DAY86	327,680
				Geomean	220,512
8	8303	GP10A	F1-WV	+DAY86	163,840
Α	8304	GP10A	F1-WV	+DAY86	163,840
8	8305	GP10A	F1-WV	+DAY86	163,840
0A	8306	GP10A	F1-WV	+DAY86	163,840
0B	8307	GP10A		+DAY86	163,840
1A	8308	GP10A	F1-WV	+DAY86	81,920
1B	8309	GP10A		+DAY86	163,840
2A	8310	GP10A		+DAY86	163,840
2B	8311	GP10A		+DAY86	163,840
3A	8312	GP10A		+DAY86	163,840
3B	8313	GP10B		+DAY86	81,920
4A	8314	GP10B		+DAY86	327,680
4B	8315	GP10B		+DAY86	327,680
5A	8316	GP10B		+DAY86	163,840
5B	8317	GP108		+DAY86	327,680
				Geomean	171,589
3A	8318	GP11	PLAGUE USP	+DAY86	
88	8319		PLAGUE USP		1,280
7A	8320		PLAGUE USP	+DAY86	640
B	8321		PLAGUE USP	+DAY86	1,280
BA .	8322		PLAGUE USP	+DAY86	640
38	8323		PLAGUE USP	+DAY86 +DAY86	1,280
A	8324		PLAGUE USP	+DA186	1,280
B	8325		PLAGUE USP	+DAY86	1,280
)A	8326		PLAGUE USP	+DAY86	640
В	8327		PLAGUE USP	+DAY86	540 1,280
_		<u> </u>	DAGOE OSI	Geomean	970
Α	8328	cosal	LHYDRO ALONE		
B	8329		LHYDRO ALONE		640 640
Ā	8331		LHYDRO ALONE		640
В	8333		LHYDRO ALONE		
IA	8335		LHYDRO ALONE		1,280
B	8336		LHYDRO ALONE		
	8337		LHYDRO ALONE		1,280
A		C1 12/	ALL THE ALUME		640
Α				Gooman	004
	E18/ 000			Geomean [861
B	F1/V POOL			Geomean [861 655,360

PROTO	OCOL: V/F	LONG	TERM		F1 antigen TITER
Plate	Serum	Group	Treatment	Bleed	IgG(H+L)
1A	8288	GP9A	F1+V_	+DAY86	NSUFFICIENT SERUI
18	8289	GP9A	F1+V	+DAY86	20,480
2A	8290		F1+V	+DAY86	NSUFFICIENT SERUI
	8291	GP9A	F1+V	+DAY86	NO SERUI
2B	8292	GP9A	F1+V	+DAY86	40,960
3A .	8293	GP9A	F1+V	+DAY86	2,580
38	8294	GP9A	F1+V	+DAY86	20,480
4A	8295	GP9A	F1+V	+DAY86	5,120
4B	8296	GP9A	F1+V	+DAY86	20,480
5A	8297	GP9A	F1+V	+DAY86	81,920
5B	8298	GP9B	F1+V	+DAY86	40,960
6A	8299		F1+V	+DAY86	2,560
6B	8300	GP98	F1+V	+DAY86	20,480
7A	8301	GP9B	F1+V	+DAY86	40,960
7B	8302	GP9B	F1+V	+DAY86	5,120
				Geomean	15,343
ва	8303	GP10A	F1-WV	+DAY86	5,120
88	8304		F1-WV	+DAY86	20,480
9A		GP10A		+DAY86	1,280
9B	8306	GP10A		+DAY86	10,240
IOA	8307	GP10A		+DAY86	10,240
108	8308	GP10A		+DAY86	2,560
I1A	8309	GP10A		+DAY86	2,560
11B	8310	GP10A		+DAY86	5,120
i2A	8311	GP10A		+DAY86	1,280
2B	8312	GP10A		+DAY86	5,120
3A	8313	GP108		+DAY86	1,280
38		GP10B		+DAY86	2,560
14A	8315	GP10B		+DAY86	5,120
14B	8316	GP10B		+DAY86	5,120
15A	8317	GP108		+DAY86	10,240
	551.7			Geomean	4,256
58	8318	6011	PLAGUE USP	+DAY86	5,120
6A	8319		PLAGUE USP	+DA186	5,120
68	8320		PLAGUE USP	+DAY86	10,240
17A	8321		PLAGUE USP	+DAY86	10,240
7B	8322		PLAGUE USP	+DAY86	10,240
8A	8323		PLAGUE USP	+DAY86	5,120
188	8324		PLAGUE USP	+DAY86	10,240
19A	8325		PLAGUE USP	+DAY86	20,480
98	8326		PLAGUE USP	+DAY86	5,120
20A	8327		PLAGUE USP	+DAY86	10,240
UA	0327	GFII	FLAGUE USF	Geomean	8,317
1					
208	8328		ALHYDRO ALONE	+DAY86	320
21A	8329		ALHYDRO ALONE	+DAY86	320
21B	8331		ALHYDRO ALONE	+DAY86	320
22A	8333		ALHYDRO ALONE	+DAY86	320
228	8335		ALHYDRO ALONE	+DAY86	320
23A	8336		ALHYDRO ALONE	+DAY86	320
238	8337	GP12	ALHYDRO ALONE	+DAY86	320
				Geomean	320

APR 96

500ml, undustrad

9AFR 96

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MAL V-YP 7F5-1-1, SUBCLONES .-10

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F1/V POOL



			Mean	1 time	time-to-death (MTD)	ath (N	(DTD)							
											MTD		Std	
Challelige	•	>	د	•	n	מ	7	æ	9	10		+/- Stdev	error	Group
group		2	ယ	4	U	6		α	ď	5			9	9
Subcutaneous:														
1 alhydrogel alone,	days	0, 30, sc									1	\rightarrow	2 0 0 0 0 0	-
Ì		6	ဝ	7	7	8	8	8	10	11	7.6	1.83/8/3	0.01202	
ogel +	io i	1-WV fu	F1-WV fusion protein day		0, 30, sc)	>	
C12 100	28	28	28	28	28	28	28	28	28	28	28			
T	10 µg Mau	ıro-V ure	a, days	0, 30, sc								1		Ì
- 1	6	28	6 28 28	28	28	28	28	28	28	28	25.8	6.95/011	2.319	
+	13.6 µg F	1-WV fu	F1-WV fusion protein day	tein day	0, 30, sc)		3	
C12 Max	၈	28	28	28	28	28	28	28	28	28	25.8	6.95/011	2.318	İ
+	27.2 µg F	1-WV fu	F1-WV fusion protein day	tein day	0, 30, sc	,					3		>	
C12 Max	28	28	28	28	28	28	28	28	28	28	2.0	C	0	١
6 alhydrogel alone,	ne, days 0,	0, 30, sc		-										İ
		ω	သ	3	3	ω	ω	5	5	თ	3.6	0.966092	0.32203	6
7 alhydrogel + 2	27.2 μg F	1-WV fu	F1-WV fusion protein day		0, 30, sc	O								
	28	28	28	28	28	28	28	28	28	28	28	0	0	!
8 alhydrogel alone,	days	,0							:					
CO92 100	ဒ	ယ	4	4	4	4	တ	6	7	7	4.8	1.549193	0.5164	∞
Aerosol:														
9 alhydrogel alone,	days	0, 30, sc												
C12 50	3	3	ယ	ω	ω	ω	ω	ω	ဒ		ဒ	0	0	9
10 alhydrogel +	13.6 μg	F1-WV fusion	usion pr	protein day	0, 30,	SC								
C12 50		28	28	28	28	28	28	28	28	28	28	0	0	10
11 alhydrogel +	1 0	μg Mauro-V υ	urea, days	0, 30,	SC									
C12 Max	ກ	28	7	28	28	28	28	28	28	28	23.6	9.287985	3.096	_
12 alhydrogel +	-	F1.WV	iusion pr	F1-WV fusion protein day	0, 30,	SC								
	13.6 µg)	3	28	28	28	28	28	28	0	0	_

MTDeath96/0515

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19	0.41833	0.83666	3.8			_			CT	4	4	3	သ	C12 Max
											č	0, 30, s	ne, days	19 alhydrogel alone, days 0, 30, sc
18	0.17496	0.46291	3.25			3	ы	ω	3	3	з	4	4	C12 Max
											30, sc	days 0,	vaccine,	18 Greer plague vaccine, days 0, 30, sc
17	0	0	28		28	28	28	28	28	28	28	28	28	C12 Max
								sc	/s 0, 30,	s V , day	Mauro's	+ 10 μg	10 μg F1	7 alhydrogel + 10 μg F1 + 10 μg Mauro's V , days 0, 30,
16	2.62561	7.87683	5.6	3	З	ω	ω	ω	3	3	3	4	28	CO92 100
											గ	0, 30, s	ne, days	16 alhydrogel alone, days 0, 30, sc
15	0	0	28	28	28	28	28	28	28	28	28	28	28	CO92 100
								SC	y 0, 30,	otein da	usion pro	=1-WV fi	13.6 µg l	15 alhydrogel + 13.6 μg F1-WV fusion protein day 0, 30, sc
74	0.11785	0.333333 0.11785	3.11111		4	3	З	ω	ယ	з	3	3	з	C12 Max
											ဂိ	0, 30, s	ne, days	14 alhydrogel alone, days 0, 30, sc
13	0	0	28	28	28	28	28	28	28	28	28	28	28	C12 Max
		A 4 A 4						SC	y 0, 30,	otein day	usion pro	-1-WV fi	27.2 μg F	13 alhydrogel + 27.2 μg F1-WV fusion protein day 0, 30, sc
	error	+/- Stdev		10	9	8	7	6	Sī	4	ယ	2	_	group
	Sto	3	MID											Challenge

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Date: 28Ju	n96		Proi	ect:	Lon	gter	m F	1-1	V, F	1+1	V C	halle	nge	, Da	y 2	16						~		-			******			com/e	approc	rysacin	er er man	
Notebook																									***	well makes to the		-	W.P-CC	المداخة والم		er breed	,	
inoculum:	Vosels	<u> </u>	aeti	e et	rein	CC	192	_																										
		ט אוו	C-C	3 31	, airi	, –	bal																~~~		~~~		-1		,_					-
floule: aer	osol				s sh	own	Dei				014	7/0/		7.0	2			Von.	lor: F	Jarle	- C	neac	un f	1000	des			Ser:	los	nale			غود سعونه	
Animal stra	in: Sw	riss V	Veb:	ster				-	_			_	at		3wks	<u>. </u>		Veri	101. r	THIR	311 3	11111	0.00		1		~-			24		1, 14	49	
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QP13A	CO92	- 1	Ш	+-	1	+		₩	┰	++	: †	+	÷	귀		1	1	11	-	\Box	71	7	71	푀	$\neg \sqcap$	1	1	\neg	\sqcap	$\overline{}$	٠,	1	l	222241257B/LT-122
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	CO92 3/20 CO92 CO92	BLAKE LINETE TOPOLE LAHINS		COXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX	XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX		×××××××××××××××××××××××××××××××××××××			ルークー				XXXXX	(XXXX) (XXXX) (XXXX) (A) (A) (A) (A) (A) (A) (A) (A) (A) (A	×××× ×××× //s	XXXXX XXXXX XXXXX XXXXX XXXXX XXXX XXXX XXXX	XXX XXXX XXXX I	000000 00000 00000 00000 00000 1		DOXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX			000 000 000 000 000 000 000 000 000 00							XXX	000	00XX 000X	2221754939/LT-138 221D5B0C51/LT-139 7 222153781B/LT-140 221B2E7B71/LT-236 221D484645LT-237 22276D3C14/LT-238 221D4C655D/LT-239 2222516E3F/LT-240 XXX XXX XXX XXX XXX XXX XXX XXX XXX X

tead animals

tead animals

fib number of dead mice with scanner

there is nitipals alive in each cage

the Chp # means the chip has fallen out

HEAD COLOR RUN 1 AFROCOL.
BUAGE RUN 2 AFROSOL

The counts were as follows:

Prespray -- $4.2 \times 10e9 (48/34/43 \text{ on } 10e7 \text{ plate})$

 AGE^{-} #1 -- 2.6 x 10e7 (18/36/25 on 10e5 plate)

#2 -- 2.2 x 10e7 (222/196/229 on 10e4 plate)

£uann

AER: IL EXPOSURE SHEET

porational Check Performed:				el. Hum.:		<u>.</u>	agent: Plays - COFA Protocol #: P.I.: LTC Andress
Stom Flow Rate: 14 Lpm	•		Timer Che Start time	ck: 1	1104	- U.S.	
nisan#: A		-	End time		1109		
ectronic Flow Meter #: Eugng					K		
Animal Sta	nt T		5 min T Wet	Rei. Hum.	Start Time	AGI#	Comments
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Active Immunization

Date:

28-Jun

LTC Anderson

Plague Agent:

CO92

Animal Model: Mouse

Wt: (Ave.) 34.7g

CO92

LD50 = 2.1E + 04

	AGI	AGI	Aerosol	WV	Inhaled Dose		Plague
Run #	cfu / ml	ctu	cfu / I	i	cfu	LD50s	Strain
1		2.60E+08	4.33E + 06	0.03	1.30E+06	61.90	CO92
,	2 20E + 07	2.20E+08	3.67E + 06	0.03	1.10E+06	52.38	CO92

	l: Long		Dave	Day +204	26JUN96	25JUN96
	21JUN96		Group	TREATMENT	F1 TITER	V TITER
بسبيد	Serum		GP13A	10F1+20V	10,240	20,480
1A		8906	GP13A	10F1+20V	1,280	40,960
1B		8907	GP13A	10F1+20V	10,240	81,920
2A		8908	GP13A	10F1+20V	10,240	81,920
2B		8909		10F1+20V	5,120	20,480
3A		8910	GP13A	10F1+20V	20,480	20,480
3B		8911	GP13A	10F1+20V	10,240	40,960
4A		8912	GP13A	10F1+20V	40,960	10,240
4B		8913	GP13A	10F1+20V	2,560	81,920
5A		8914	GP13A	10F1+20V	10,240	81,920
5B		8915	GP13A	10F1+20V	5,120	20,480
6A		8916	GP13B	10F1+20V	20,480	81,920
6B		8917	GP13B	10F1+20V	2,560	40,960
7A		8918	GP13B	10F1+20V	20,480	163,840
7B	<u> </u>	8919	GP13B	10F1+20V		81,920
8A		8920	GP13B	30ugF1-V		81,920
8B	1	8921	GP14A	30ugF1-V		81,920
9A		8922	GP14A			40,960
9B		8923	GP14A	30ugF1-V		40,960
10A		8924	GP14A			
10B		8925	GP14A			81,920
11A		8926	GP14A			
11B		8927	GP14A			
12A		8928	GP14A			
12B		8929	GP14A			
13A		8930	GP14A			
14A		8931	GP 14E	30ugF1-\	/ 10,240 / 20,480	
14B		8932	GP14E			
15A		8933	GP148			
15B		8934	GP14E			
16A		8935	GP14F			
16B		8936	GP1	PlagueUS		
17A		8937	GP1	PlagueUS		
18A		8938	GP1	PlagueUS		
18B		8939	GP1	5 PlagueUS		
19A		8940	GP1	5 Plag ueUS	P 2,560	
19B		8941	GP1	5 PlagueUS	P 5,120	
20A		8942	GP1	5 PlagueUS		
20B		8943	GP1	5 PlagueUS	P 20,48	
21A		8944	GP1	5 PlagueUS	P 40,96	
21B		8945	GP1		P 5,12	
		8946	GP1		i e 64	0 2,560
22A		8947	GP1			
228		8948	GP1			
23A		8949	GP1			0 1,28
23E		8950	GP1			
244			GP1			0 64
24E		8951 8952	GP1			
25/			GP1			
258		8953	GP.	-		
26/		8954	GP.			
26		8955		101 712 010	327,€8	
	F1/V		POOL			20 2,56

GEOMEAN:	Group	TREATMENT	हा ग्री	V TITER
GEUWEAN	GP13A/B		8.:.12	44,926
	GP14A/B			85,794
	GP15	71.00		2,229
	GF 13		:20	1,194

for plague shelling of 5 of 16

74 495 C12

lets from bur Nechon

1 1 ac
8 11196

Sky -

plague-challenge.sc 7/5/96

7/5/96

P.I. = LTC George Anderson 40 mice, C092 - 100 LD50s 30 mice, C12 - 100 LD50s

Parenteral challenge of mice with C092/M.S. and C12/M.S.

- Streak 1 slant each with the Master Seed of C092 and C12. Incubate 2 days at room temperature.
- 2. Harvest by suspending in 4-5 mls of HIB.
- 3. Read OD620 of a 1/10 dilution.
- 4. Adjust to OD 1.0

7/5/96:

Adjusted ODs and read final ODs on 1/2 dilutions:

Final OD = <u>1.064</u>, for C092 " - <u>1.10</u>, for C12

C092/M.S.:

1. Prepare dose

5.0 -7.5x10e2/ml:

- (1) Add <u>0.2 ml</u> OD 1.0 to <u>1.8 mls</u> HIB.
- (2) Add 0.2 ml (1) to 1.8 mls HIB.
- (3) Add 0.5 ml of (2) to 4.5 mls HIB.
- (4) Add 0.5 ml of (3) to 4.5 mls HIB.
- (5) Add <u>0.5 ml</u> of (4) to <u>4.5 mls</u> HIB.
- (6) Add 4.0 ml of (5) to 36 mls HIB - Pipet 10 mls into each of 3 tubes: mice INOCULUM: 1 x 10e3/ml: ~200 cfu/dose
- 2. Plating: The sample will be diluted in HIB and plated on SBAP:

Total No.

<u>suspension Conc./ID dilution no. plates plates</u>
mice Inoculum 5x10e2/ml undil, 10-1 5 each 10

RESULTS:

7/5/96 doses: 1.4 x 10e3/ml, 280 cfu/dose (140 LD50s)

7/12/96 doses: 6.5 x 10e2/ml. 130 cfu/dose (72 LD50s)
7/18/96 doses: x 10e2/ml. cfu/dose (LD50s) - Correled

C12/M.S.:

1. Adjust slant suspension to OD620 = $\underline{1.0}$. Prepare dose

2.3 x10e3/ml:

1

more	(1) Add <u>0</u>	.2 ml OD 1.0 to 1.8	mls HIB	
34,2	(3) Add 0	5 ml of (0)	HIB.	
	(4) Add <u>0</u>	.5 ml of (3) to 4.5 m	nis HiB. nis HiB	
The count	(5) Add <u>1</u> (6) Add 6	0 ml of (4) to 9.0	mls HIB.	
Pres AGE		- III or (2) to 18	mis HiB (1/4)	
#1			NOCULUM, C12-100 sc L mls into each of 2 tu	
#2			1 x 10e3/ml: ~200 c	fu/dose
	2. Plating: The Inc	Culum will be an .		
	Suspansia	diuted	I in HIB and plated on SBAP	
	suspension C12 Inoculum	Conc./ID	Total No.	10. plates plates
		2.3x10e3/ml	unun.	
			10-1 10-2 5	
Amnia	RESULTS:			TOTAL-15
Sup. 611	7/5/96 doses: 7/12/96 doses:	3.36 x 10e3/ml.	6.7 x 10e2 cfu/dose	
Syste	7/18/96 doses:	3.0 X 10e3/ml. X 10e3/ml.	3.8 X 1082 cfu/dose	/ 62 7 LDTS
Com		- 1000/IIII:	x 10e2 cfu/dose	(LD50s) - comules
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AIBS PEER REVIEW TO USAMRMC Medical Biological Defense Research Program ON PLAGUE

REVIEW PANEL

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EXECUTIVE BECRETARIAT

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APPROYED:

Tom Schwan, PhD

KATHLEEN MCDONONEN PLD

DATE: March 12,1996

INTRODUCTION

AIBS was requested by US Army Medical Research and Development Command (USAMRDC) to convene a review Panel to provide an assessment of the scientific merit of the Medical Biological Defense Research Program (MBDRP) on Plague. It was requested that the three scientific reviewers have a collective knowledge of the following subject areas: Yersinia pestis, Vaccine Production, Molecular Genetics and FDA requirements for a vaccine. Such a panel was convened and provided with documentation by USAMRDC to read prior to the conference. This consisted of abstracts prepared by the individual investigators who form the MBDRP on Plague (see Appendix 1.)

CHARGE TO PANEL

Three scientific reviewers were asked to evaluate the MBDRP on Plague. They independently reviewed material provided by USAMRDC and attended a conference on the subject matter. They were asked to judge the scientific merits of the Program.

The reviewers, individually, provided comments to AIBS, who in turn compiled this written report summarizing these comments and the discussions at the conference. The Chairman of the Review Panel read and approved the report prior to its submission to USAMRDC.

PRESENTATION SUMMARIES

The conference comprised presentations by each of the following investigators. Abstracts were provided for by each and are attached as Appendix 1.

COL ARTHUR FRIEDLANDER
Overview of plague program

COL RUSSELL BYRNE
Antibiotic treatment of experimental pneumonic plague

DR. PATRICIA WORSHAM, DR. M. LOUISE PITT, LTC KELLY DAVIS F1 is not a required virulence factor for the mouse or non-human primate

MAJ GERALD P. ANDREWS, LTC GEORGE J. ANDERSON, JR. Protective efficacy of active immunization with purified F1 from Yersinia pestis and an Escherichia coli recombinant strain against lethal parenteral and respiratory plague challenge

DR. PATRICIA WORSHAM
Studies on the role of the pigmentation locus in the pathogenesis of Y. pestis

DR. SUSAN L. WELKOS, LTC KELLY J. DAVIS

Analysis of the role of pPst encoded genes in pathogenesis of infection by Y. pestis

DR. ALAN SAMPLE

Plasminogen activator protease degrades proinflammatory cytokines

MAJ GERALD P. ANDREWS, DR. SUSAN STRALEY, DR. ALAN SAMPLE, MAJ GERALD P. ANDREWS

Cloning, Expression, Purification, and Protective Efficacy of Yops and pH 6 antigen

LTC GEORGE J. ANDERSON, JR., DR. DAVID HEATH Cloning, expression, and protective efficacy of V antigen

LTC GEORGE J. ANDERSON, JR.

Cloning, expression, and protective efficacy of F1-V fusion protein

DR. JEFFREY PULLEN

Determination of important B and T-lymphocyte epitopes in the F1 and V antigen proteins of Yersinia pestis

COL ARTHUR FRIEDLANDER
Overview of future plans

SUMMARY EVALUATIONS OF THE RESEARCH AREAS

The review panel read the abstracts provided by the investigators prior to the meeting on February 15, 1996, and listened to presentations by each of the investigators at the meeting. The following comments include recommendations to individual investigators, and are intended to be constructive. Certain points apply to more than one project, or even to the program as a whole, and hence may appear repetitive. Also, the reviewers recognize that some of their recommendations may be affected by programmatic decisions that are beyond the control of the immediate Program staff and thus may not prove to be possible.

COL ARTHUR FRIEDLANDER Overview of plague program

The USAMRMC Plague Research Program's primary objective is to develop a vaccine that will protect military personnel if exposed to an aerosol attack of Yersinia pestis, the causative agent of plague. Given that the currently available vaccine (USP) protects primarily through anti-F1 antibody, that this vaccine offers very poor protection from primary pneumonic plague, and that F1⁻ strains are highly virulent, there clearly is a need for a new, more protective vaccine. Once developed, the general population living in areas endemic for plague would also benefit from such a vaccine.

Most of the projects presented as separate studies and presentations clearly meet the program's primary objective. Part of the rationale for the approach taken is that an aerosol attack of *Yersinia pestis* might include strains that do not produce the F1 capsular antigen. Given that the current vaccine (USP) stimulates primarily antibodies

AIBS PEER REVIEW TO USAMRDC MEDICAL BIOLOGICAL DEFENSE RESEARCH PROGRAM ON PLAGUE

TIME:

February 15, 1996, 8.00am to 5.00 pm

LOCATION:

US Army Medical Research Institute of Infectious Diseases, Conference Room, Fort Detrick, Frederick, MD

EXECUTIVE SUMMARY

Overall, the program has made very significant and impressive advances in only a few years towards the development of a new vaccine, and Dr. Friedlander and his entire team of investigators can be proud of their accomplishments to date. They clearly have a very viable, sound program with a good team of investigators that is focused with high potential to succeed. It is hoped that the administration will continue to support this effort and provide the group with the resources and time necessary to complete their task. The investigators clearly considered the recommendations of the previous reviewers and incorporated several of the suggestions into their program.

The team has invested significant effort in examining numerous virulence determinants of Yersinia pestis for their ability to stimulate protection through immunization. The F1 capsular antigen and the V antigen have been shown by investigators in other laboratories to be good candidates for inclusion in a new multivalent subunit vaccine. The team at USAMRMC has confirmed the protective value of these two antigens. However, realizing that F1 and V antigens might not be sufficient for full protection against all virulent strains of Y. pestis, the group has worked through an impressive list of additional candidates. The only other antigen that offered significant protection was YopD, although protection was only observed when mice were challenged with the F1 strain. Passive immunizations with anti-F1 and anti-YopM antisera deserve further attention. Combined antibiotic treatment and immunization might increase the survival of animals challenged by aerosol.

The team appears to make use of mice and nonhuman primates as excellent animal models for both their parenteral and aerosol challenge experiments. The current vaccine study protocols for test challenges are very good.

The development of *in vitro* correlates of immunity should be a high priority of the program. It is currently the weakest portion of the future plans. As discussed with the investigators, the assumption that protection is solely antibody-mediated has potential difficulties. Before continuing studies to map active B cell epitopes, the investigators need to determine the role of T cells in immunity to plague.

examined for Y. pestis in the LD50 studies and survivors were examined for clearance of the organisms to determine the full level of protection provided by vaccination.

In the first study, the V antigen was examined for its ability to generate a protective immune response in mice challenged by parenteral subcutaneous or aerosol challenge with either the F1+ or F1- isogenic strains of *Y. pestis*. Recombinant V antigen was cloned and expressed in two fusion/expression systems and used with an adjuvant approved for human use (Alhydrogel). Both preparations of rV antigen were administered twice and provided very good protection in mice challenged by both routes and both strains. This is an excellent study and identifies (as another laboratory has demonstrated independently) the V antigen as an excellent candidate immunogen to include in a vaccine to protect from aerosol infections with either F1+ or F1- strains. These studies are critical to the program's objective and provides some quite exciting results.

The second study extends the work on the V antigen of Y. pestis by examining protection following a single dose of 10 µg (the previous study used two immunizations prior to challenge). Mice were subsequently challenged by aerosol exposure to either low and high doses of the F1+ or F1- strain. Protection ranged from 70% to 78% survival in these mice, demonstrating that a single immunization could afford significant protection from an aerosol route of infection. However, the schedule including a primary immunization followed by a single boost afforded 20% to 30% greater protection (previous report). While it is of interest what level of protection results from a single dose, future work with nonhuman primates will likely confirm what we know about many other bacterial vaccines, i.e., better protection results with boosts following the primary immunization.

Two areas need to be addressed in future work on the V antigen. The studies presented used the V antigen tagged with histidine from the pET vector. If this antigen is to be used in humans, a method for the efficient removal of the his-tag is needed. Identifying the active sites on the V antigen responsible for protective immunity as well as potential negative biological activities, such as immune suppression, may be required for this antigen to be safe. The group might also consider examining how long protective immunity lasts following vaccination with the V antigen. Some of these issues were addressed by Dr. Friedlander in his closing remarks.

LTC GEORGE J. ANDERSON, JR. Cloning, expression, and protective efficacy of F1-V fusion protein (abstract 17)

Prior studies have confirmed the potential for both F1 and V antigen to protect mice from Y. pestis by both parenteral and aerosol routes. In this study a construct was made containing the F1 and V antigen genes for expression of a fusion protein. When the F1-V fusion protein was used for immunization, mice were protected when challenged by needle or aerosol with either the F1 positive or F1 negative strain of Y. pestis. Poorer protection resulted when only a portion of the V antigen was expressed as a fusion protein with F1. This work is quite clever and interesting, and advances the program's effort towards the development of a multivalent vaccine. The attempt to make fusions of these two antigens also demonstrates an advance towards reducing

the steps required for making and purifying antigens for the vaccine. The investigators are also testing longer term antibody responses and how long protection lasts (a concern raised from the previous studies with the V antigen alone). Antibody responses to the F1 and V antigen components of the fusion protein were also examined. Both F1 and the V antigen have been shown by other workers to be protective and now the group at USAMRMC has shown that rF1 and rV are the best candidates identified to date for a new plague vaccine.

Again, this fusion protein has a histidine tag, which will need to be removed prior to its use in humans.

DR. JEFFREY PULLEN

Determination of important B and T-lymphocyte epitopes in the F1 and V antigen proteins of *Yersinia pestis* (abstract 18)

This study attempts to identify important B and T cell epitopes within both the F1 and V antigens, however only B cells were addressed in the presentation. Identifying the functional epitopes in these proteins is important both to an understanding of the protective mechanisms stimulated by these two immunogens, and for assessing the potential of using synthetic peptides rather than entire recombinant proteins in a vaccine. This study is an important part of fulfilling the long-term objective of developing a useful vaccine. However, the usefulness of the current appproach should be carefully reconsidered.

The use of short peptides to generate antibodies without conjugation to carrier molecules has, in general, not been very successful. Although it is sometimes possible to generate antibodies against short peptides, it is unlikely that the response will be protective without some T cell involvement. The investigators' initial experiments showed that peptides generated from the region of the protein known to be antigenic failed to generate a protective response despite generating significant antibody production. These results should have alerted them to the problems inherent in this approach. Instead, the investigators expanded their studies in response to these findings by making and testing additional peptides covering the whole of V antigen and F1 protein. This was a lot of work, using a lot of mice, that generated very little useful information. A simpler and more direct approach to begin mapping the reactive epitopes in these immunogens is to screen the overlapping peptides in vitro using antisera from animals or humans that have either had infections with Y. pestis or been immunized with native F1 and/or V antigen. Another concern is that in the future goals, it was stated that the response to the peptides, rather than to the native antigen will be tested to better determine the response. However, since the goal is to get protective antibodies, it seems that the reponse to native antigen, which is what the animals will see in an actual infection, is what should be measured.

It is also important for the investigators to determine the nature of a protective immune response to *Y. pestis* infection before restricting their focus and undertaking such labor-intensive studies to define only B cell epitopes. Antibody reactivity does not assure protection, and with some pathogens high antibody titers have even been correlated with disease progression. In addition, non-F1 antigens may evoke a

COMBINED RECOMMENDATIONS AND CONCLUSIONS

The USAMRMC's program to develop a new subunit vaccine for pneumonic plague has been very productive and has made significant advances towards this objective. The leader and research team are highly skilled, competent investigators and, with continued support, it is anticipated that a new vaccine for human trials is only a few years away. The investigators have used very effective immunization and challenge protocols to test immunogens in both mice and nonhuman primates for protection against plague following either parenteral or aerosol exposures to *Yersinia pestis*. Having the facilities to safely execute aerosol transmission studies is a critical component of this program. The team has confirmed and extended the data supporting the potential for both recombinant F1 and V antigens to afford significant protection. The work using the F1-V antigen fusion protein is exciting and represents a significant advance made by this team.

The team has examined numerous other antigens for identifying additional protective immunogens, especially for challenge with strains of Y. pestis lacking the F1 antigen. For such isolates, the V antigen and possibly YopD are the only useful candidates identified to date. The addition of one more antigen would likely solve the problem of non-responders, as well as strengthen the response in all individuals. The choice of antigens being tested for potential vaccine components appears somewhat random. These studies could be focused better by determining what proteins induce an immune response, thereby demonstrating which determinants are most likely being seen by the immune system. Although it is not possible to predict in advance which antigens are protective, the search could have been directed more towards antigens known to induce an antibody response in infected human patients and laboratory infected animals. Additional focus on the basis of immunity to plaque challenge is also recommended. The investigators are also aware of the immunosuppressive effects of V antigen, and plan to examine the mechanisms involved. These types of studies should allow the team to "fine tune" the V antigen to increase its efficacy and safety as a vaccine component.

The development of *in vitro* correlates of immunity should be a high priority of the program and is currently the weakest area of the future plans. As discussed with the investigators, the assumption that protection is solely mediated by antibody has potential difficulties. Before continuing studies to determine important B cell epitopes, the role of T cells needs to be addressed in collaboration with immunologists. There are standard methods, such as adoptive transfer, to determine if T cells protect against challenge with *Y. pestis*. There are also *in vitro* techniques to determine if T cells taken from an immunized animal proliferate in response to specific antigens. The studies using synthetic peptides have potential, but this work needs to be done with conjugated peptides. Alternatively, peptides could be attached to larger inert particles that could be taken up by B cells or macrophages that then present the antigen on class II MHC molecules on their surface. Epitope mapping of the F1 and V antigen peptides using immune sera from natural infections would have been an appropriate first step.

APPENDICES

APPENDIX 1: AGENDA

APPENDIX 2: ABSTRACTS

REVIEW OF PLAGUE RESEARCH PROGRAM

USAMRIID

15 FEBRUARY 1996

0815-0830 Welcome and introduction COL David Franz, DVM, Ph.D.

0830-0900 Overview of Plague Program

COL Arthur M. Friedlander, M.D.

Treatment

0900-0930 Antibiotic treatment of experimental pneumonic plague COL Russell Byrne, M.D.

Role of F1 Capsule in Pathogenesis and Immunity

0930-1000 Protective efficacy of active immunization with purified F1 from Yersinia pestis and an Escherichia coli recombinant strain against lethal parenteral and respiratory plague challenge

MAJ Gerard P. Andrews, Ph.D.

1000-1015 Coffee Break

1015-1100 F1 capsule is not a required virulence factor for the mouse or non-human primate

Patricia L. Worsham, Ph.D. M. Louise Pitt, Ph.D. LTC Kelly J. Davis, DVM

Role of Non-F1 Proteins in Pathogenesis and Immunity

1100-1130 Studies on the role of the pigmentation locus in the pathogenesis of *Y. pestis*Patricia L. Worsham, Ph.D.

1130-1300 Lunch

- 1300-1320 Analysis of the role of pPst encoded genes in pathogenesis of infection by *Y. pestis*Susan L. Welkos, Ph.D.
- 1320-1335 Plasminogen activator protease degrades proinflammatory cytokines

 Allen Sample, Ph.D.
- 1335-1405 Cloning, expression, and protective efficacy of Yops and pH 6 antigen
 MAJ Gerard Andrews, Ph.D.
- 1405-1420 Cloning, expression, and protective efficacy of V antigen LTC George J. Anderson, Jr., Ph.D.
- 1420-1435 Cloning, expression, and protective efficacy of F1-V fusion protein

 LTC George J. Anderson, Jr., Ph.D.
- 1435-1450 Determination of important B and T-lymphocyte epitopes in the F1 and V antigen proteins of *Y. pestis*Jeffrey Pullen, Ph.D.
- 1450-1515 Overview of future plans COL Arthur M. Friedlander, M.D.

Recombinant F1-V (rF1-V) Fusion Protein Protects against Lethal Wildtype Yersinia pestis in a Mouse Model

DAVID G. HEATH, GEORGE W. ANDERSON, JR., CHRISTOPHER BOLT, SUSAN L. WELKOS, PATRICIA L. WORSHAM, AND ARTHUR M. FRIEDLANDER Bacteriology Division, U.S. Army Medical Research Institute of Infectious Diseases, Ft. Detrick, MD.

The virulence of F1- strains and their occurence in nature imply that F1 immunogen will not be sufficient for an optimal new A fusion protein has the theoretical possibility plague vaccine. of simplifying and reducing the cost of production of multiple antigens in addition to stabilizing the protein. For these reasons, we developed a fusion protein consisting of both the F1 and V antigens (1). The first fusion protein made consisted of F1 fused with residues 168-175 of the V antigen, a segment which previous studies suggested to contain a protective epitope. This fusion protein was used with the adjuvant alhydrogel (aluminum hydroxide) to immunize female Swiss Webster (Hsd:ND4) mice subcutaneously (s.c.) on days 0 and 30 followed by a s.c. or aerosol challenge with either the F1- C12 strain (LD50 = 9.1 CFU, s.c.; LD50 = 1.1 x 105 CFU, aerosol route) or the wild-type F1+ CO92 (LD50 = 1.9 CFU, s.c. route; 2.1 x 104 CFU, aerosol route) strain of Y. pestis. Endotoxin had been removed from the preparations prior to immunization, so that this would not be a confounding factor.

When 18.5 μg of the F1-V168-275 fusion protein was used to immunize mice, there was 90% survival (9/10) when challenged s.c. with 63 LD50 of the F1+ CO92 strain. The positive control was a group of mice immunized with 10 μg of rF1 which is equivalent to the F1 content of the F1-V168-275 protein. The rF1 control resulted in 100% (10/10) protection. The F1-ELISA IgG titers were the same (1:81920). All mice in alhydrogel control group died (0/9; MTD \pm SD, 5.2 \pm 1.0). When the F1-V168-275 immunized mice were challenged with 104 LD50 by the aerosol route, protection was 80% (8/10; MTD \pm SD, 20.3 \pm 7.1) compared to 0% for the control group (0/10; MTD \pm SD, 3.1 \pm 0.3; 80-104 LD50). The group immunized with rF1 resulted in 70% protection (7/10; MTD \pm SD, 9.0 \pm 1.0) when challenged with 80 LD50. The addition of part of the V protein onto the F1 protein did not appear to effect its antigenicity.

The F1- strain, C12, was used to test the ability of the partial V portion of the F1-V168-275 protein to protect mice against a lethal challenge. Here 27 μg of the F1-V168-275 fusion protein was used, which is equivalent to 10 μg of the V protein

known to be protective. A s.c. challenge dose of 55 LD50 resulted in 30% survival $(3/10, MTD \pm SD, 9.4 \pm 7.0)$. All of the controls died (0/10, MTD \pm SD, 10.8 \pm 4.8). While there was some protection, there was no increase in the MTD. There was a good V-ELISA antibody response to the F1-V168-275 (1:163840). this response was not sufficient, another group was immunized with 27 μg, but with complete Freund's adjuvant (CFA). In this case, protection was only 20% (2/10, MTD \pm SD, 9.1 \pm 3.2), while 10 μ g of rV in CFA resulted in 100% protection. The V-ELISA titer when CFA was used was 1:1310720 for F1-V168-275 and rV. A 10-fold increase in the V-antibody titer did not have any effect on protection and the V-ELISA titer was not indicative of protection. When a group of F1-V168-275 mice were challenged with 95 LD50, C12, by the aerosol route, no mice survived (0/10, MTD \pm SD, 3.5 \pm 0.5) All of the alhydrogel control group died (0/10, MTD \pm SD, 3.4 ± 0.5). In other experiments, rV itself gave 80-90% protection against an aerosol challenge.

These results demonstrated the feasibility of making a F1-V fusion protein. The efficacy of F1 was not altered by making a fusion protein. However, while the V168-275 protein portion of the fusion protein was antigenic, it was not immunogenic. This caused us to address the question as to whether the entire V protein could be fused to F1 and whether it would be immunogenic.

Using a fusion protein which combines the whole F1 and the whole V protein (rF1-V) to immunize mice on days 0 and 30 increased the protection afforded by the V portion of the fusion protein. When 13.6 ug of rF1-V was used to immunize mice, there was 100% (10/10) protection against a s.c. challenge of 57 LD50 and 90% (9/10) protection against 1.1 x 106 LD50 C12 strain. micrograms (10 μg) of rV also gave 90% (9/10) protection against 1.1 x 106 LD50, C12 strain. All of the alhydrogel control group died (0/10, MTD \pm SD, 6.0 \pm 0.0). The rF1-V protein also offered protection against an aerosol challenge. The same immunization schedule resulted in 100% (10/10) when mice were challenge with 546-636 LD50, C12 strain on day 73 postimmunization. When mice immunized with the rF1-V fusion protein were challenged with 762 LD50 of the F1+, CO92 strain by the aerosol route, 100% (10/10) of the mice survived. The F1-V fusion protein was able to protect mice from a significant aerosol challenge from either a F1+ or F1-This protection is better than the lethal strain of Y. pestis. protection afforded by the current Plague Vaccine USP. which were immunized on day 0 and 30 with 0.2 ml of the current vaccine and challenge by the aerosol route on day 73 postimmunization with 546-636 LD50, C12 strain, all of the mice died (0/8, MTD \pm SD, 3.3 \pm 0.5). The V-ELISA titer to the Plague

Vaccine USP was <1:640.

The recombinant rF1-V fusion protein was produced in E. coli and contained a polyhistidine tag which aids in the purification of the fusion protein. While this protein has been shown to be highly efficacious in the mouse model, it remains to be seen whether this level of protection will be seen the in the non-human primate model. Further, the regulatory issue of whether a histidine tagged protein will be acceptable to the Food and Drug Administration needs to be resolved.

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